

Burden of highly resistant microorganisms in a Dutch intensive care unit

H. Aardema^{1*}, J.P. Arends², A.M.G.A. de Smet¹, J.G. Zijlstra¹

Departments of ¹Critical Care, ²Medical Microbiology, University Medical Center Groningen, the Netherlands, *corresponding author: tel.: +31(0)50-3616161, fax: +31(0)50-3613216, email: h.aardema@umcg.nl

ABSTRACT

Background: The occurrence of highly resistant microorganisms (HRMOs) is a major threat to critical care patients, leading to worse outcomes, need for isolation measures, and demand for second-line or rescue antibiotics. The aim of this study was to quantify the burden of HRMOs in an intensive care unit (ICU) for adult patients in a university hospital in the Netherlands. We evaluated local distribution of different HRMO categories and proportion of ICU-imported versus ICU-acquired HRMOs. Outcome of HRMO-positive patients versus controls was compared.

Methods: In this prospective single-centre study, culture results of all ICU patients during a four-month period were recorded, as well as APACHE scores, ICU mortality and length of stay (LOS) in the ICU.

Results: 58 of 962 (6.0%) patients were HRMO positive during ICU stay. The majority (60%) of those patients were HRMO positive on ICU admission. HRMO-positive patients had significantly higher APACHE scores, longer LOS and higher mortality compared with controls.

Conclusions: Our study suggests that a large part of antibiotic resistance in the ICU is imported. This underscores the importance of a robust surveillance and infection control program throughout the hospital, and implies that better recognition of those at risk for HRMO carriage before ICU admission may be worthwhile. Only a small minority of patients with HRMO at admission did not have any known risk factors for HRMO.

KEYWORDS

Critical care, highly resistant microorganisms, HRMO, length of stay, surveillance, isolation

INTRODUCTION

Antibiotic resistance in the critical care population is an ever-increasing problem.¹ The high use of antimicrobial therapy in the intensive care unit (ICU),² the large number of invasive procedures, the density of a susceptible patient population, the severity of underlying illness, and flaws in infection control measures are all contributing factors resulting in ICUs as 'epicentres' of antimicrobial resistance in hospitals.^{3,4} ICUs are considered generators of antimicrobial resistance.³ In addition to acquisition of HRMOs in the ICU, part of the resistance problem is imported to the ICU through already colonised or infected patients admitted from other hospitals, general wards, or from the community.⁵

This continuous and rising threat of antimicrobial resistance is of relevance considering the outcome in patients infected with resistant rather than susceptible microorganisms is worse.⁶ Measures to prevent cross-contamination include surveillance, barrier precautions and antibiotic stewardship. All preventive measures are labour intensive, costly and some are patient unfriendly. Resistance to first-line antibiotics urges the use of 'rescue' or second-line antibiotics with little hope of new effective alternatives in the near future.⁷

The incidence and characteristics of resistance can vary widely depending on local circumstances. According to European surveillance data, the Netherlands, along with Scandinavian countries, is considered an area with low incidence of antimicrobial resistance for Gram-positive as well as Gram-negative bacteria.⁸ However, even in the Netherlands, prevalence of antimicrobial resistance in the community is not negligible, and is emerging.⁹⁻¹¹

We evaluated the incidence of HRMOs in our ICU to quantify the total burden of HRMOs, to clarify the local distribution of different categories of HRMOs and the proportion of imported versus acquired HRMOs in our

ICU. Furthermore, we evaluated the outcome of patients affected by colonisation or infection with any HRMO vs. controls in terms of ICU mortality and length of stay (LOS) in the ICU. Finally, we wanted to characterise this subpopulation colonised or infected with an HRMO to enable better *a priori* recognition of affected patients, thus rendering better opportunities for adequate treatment and infection control.

MATERIALS AND METHODS

This is a single-centre study involving prospective data collection from 1 October 2009 to 31 January 2010 in an academic teaching hospital with 40 critical care beds distributed over 4 units (medical, cardiothoracic, neurological, and general surgery). All four units consist of a large multi-bed floor combined with a few rooms for isolation. On the floors, standard hygienic procedures are maintained. Annually, between 2500 and 3000 patients are admitted. All patients admitted in the four-month study period were included for analysis. The study was approved by the ethics committee and the requirement of informed consent was waived. We recorded baseline characteristics including sex, age, referring speciality, unit of admission, APACHE II score, date and source of admission (emergency department or general ward vs. other hospital). APACHE II score was not recorded for cardiosurgical patients because this score is not validated for this subgroup. Of note, patients from various sources were admitted to the four separate units, e.g. the cardiothoracic unit did not only admit cardiothoracic surgery patients but other patients as well. Patients were considered referred from another hospital if they were transferred either directly to the ICU or indirectly through another ward in our hospital. The vast majority of this last group were admitted to the general ward for less than a week before admission to our ICU.

All cultures taken either by indication or in the context of our structured surveillance program were evaluated. Surveillance screening included cultures from throat, nose, rectum, sputum and urine on admission, followed by cultures from throat, nose, rectum and sputum on day four and twice weekly thereafter during the stay in the ICU. Surveillance cultures were obtained from those patients with an anticipated stay of 48 hours or more on the day of admission. Patients referred from elsewhere were included in surveillance screening on the day of admission regardless of anticipated or actual length of stay.

Culture results were retrieved from the database of the department of medical microbiology. Susceptibility testing was done according to European guidelines (European Committee on Antimicrobial Susceptibility Testing, EUCAST).

HRMOs were defined by criteria issued by the Dutch Working Party on Infection Prevention (*table 1*).¹² All patients colonised or infected with an HRMO were placed in full contact isolation, as dictated by our protocol for infection prevention. A patient could be included only once in the study group; subsequent readmissions of the same patient were excluded from the study group but were analysed nevertheless. Only the first positive culture for any HRMO in an individual was recorded; subsequent cultures with the same organism were regarded as the same event. Different species of HRMOs within one patient were recorded as separate events. No distinction was made between either colonisation or infection with an HRMO. Of the patients with an HRMO, further details such as antibiotic use during the hospital stay and medical history were retrieved from the patient's file. Other outcome measures for the entire study population included ICU mortality and LOS on the ICU.

We tried to identify clusters of HRMOs by analysing whether identical species of HRMOs were cultured in different patients during their ICU stay on the same unit. Statistical methods included the χ^2 -test, Fisher's exact test, Student's t-test, Mann-Whitney U-test and Wilcoxon rank test, where appropriate, using Minitab® Release 14.1 and Graph Pad Prism (Prism 5 for Windows, version 5.04, Nov 6 2010) software.

RESULTS

A total of 1061 admissions were recorded, 91 of which were re-admissions within the study period; hence 962 admitted patients were included in the study population (*table 2*). Baseline characteristics are presented in *table 2 and 3*. In 58 (6.0%) patients an HRMO was found (in total 60 HRMOs; two patients had two different HRMOs). For distribution of HRMO species we refer to *table 4*. The distribution of these 58 patients according to unit and patient category is depicted in *table 2*.

Of 232 patients (24.1%) referred from another hospital, 16 patients were colonised with an HRMO during their stay in our ICU (6.9%), compared with 42 out of 730 patients (5.8%) referred from our hospital ($p = 0.52$).

In those patients with any HRMO ($n = 58$), 47 patients (82.8%) were found to have an HRMO within the first three days of ICU stay. Of these, 11 (23.4%) were referred from another hospital: 36 (76.6%) were admitted from a general ward of this hospital or from the emergency department.

Of those not referred from elsewhere and found positive for an HRMO within three days ($n = 36$), 32 patients (88.9%) had one or more comorbid conditions (*table 4*). In this group of 36 patients, 27 patients (75.0%) had been admitted in the three months preceding current admission

Table 1. Definition of highly resistant microorganisms (HRMOs)¹²

	ESBL	Quino- lones	Amino- -glycosides	Carba- penems	Co-trimo- xazole	Ceftazi- dime	Pipera- cillin	Penicil- lins	Glyco- peptides	Oxacillin	Methi- cillin
Enterobacteriaceae											
<i>E. coli</i>	A	B	B	A	--	--	--	--	--	--	--
<i>Klebsiella</i> spp	A	B	B	A	--	--	--	--	--	--	--
Other	A	B	B	A	B	--	--	--	--	--	--
Non-fermenting gram-negative											
<i>Acinetobacter</i> spp.	--	B	B	A	--	B	--	--	--	--	--
<i>Stenotrophomonas</i> spp.					A		--	--	--	--	--
Other		C	C	C		C	C	--	--	--	--
Gram-positive											
<i>S. pneumoniae</i>	--	--	--	--	--	--	--	A	A	--	--
<i>Enterococcus</i> spp.	--	--	--	--	--	--	--	B	B	--	--
<i>S. aureus</i>	--	--	--	--	--		--	--	--	A	A

Resistance to one antibacterial agent in category A, to \geq two in category B, or \geq three in category C required to define microorganism as highly resistant microorganism (HRMO). ESBL = extended beta-lactamase (resistance to any third-generation cephalosporin used as proxy in *E. coli*, *Klebsiella* spp., and *Proteus* spp.)

Table 2. Patient characteristics

	Total	Without HRMO	With HRMO	P
Study population, n	962	904	58	
Male n (%)	595 (61.9%)	556 (61.5%)	39 (67.2%)	0.38 (NS)
Age, years, median (range)	62 (12-91)	63 (12-91)	58 (16-82)	0.22 (NS)
APACHE II score, median (range)**(n) • APACHE II > 20 (n = 101) • APACHE II \leq 20 (n = 431)	13 (2 – 52) (532)	13 (2-44) (488) 83 (17.0% of 488) 405	18 (2-52) (44) 18 (40.9% of 44) 26	< 0.001 < 0.001
Unit, n (%) • Cardiopulmonary unit • Surgical unit • Medical unit • Neurosurgical unit	390 (40.5%) 238 (24.7%) 181 (18.8%) 153 (15.9%)	381 (42.1%) 211 (23.3%) 168 (18.6%) 144 (15.9%)	9 (15.5%) 27 (46.6%) 13 (22.4%) 9 (15.5%)	< 0.001
Patient category, n (% of total) • Cardiopulmonary surgery • Medical • Surgical • Neurosurgical • Trauma • Neurological • Gynaecological	405 (42.1%) 208 (21.6%) 181 (18.8%) 103 (10.7%) 44 (4.6%) 17 (1.8%) 4 (0.4%)	392 (43.4%) 186 (20.6%) 167 (18.5%) 101 (11.2%) 38 (4.2%) 16 (1.8%) 4 (0.4%)	13 (22.4%) 22 (37.9%) 14 (24.1%) 2 (3.4%) 6 (10.3%) 1 (1.7%) 0 (0%)	

**APACHE II-score available for 532 (95.5%) of non-cardiosurgical patients; HRMO = highly resistant microorganism.

Table 3. Patient characteristics and outcome

	Total	Without HRMO	With HRMO	P
Study population, n (%)	962	904 (94.0%)	58 (6.0%)	
Referral from other hospital, patients, n (%)	232 (of 962; 24.1%)	216 (of 904; 23.9%)	16 (of 58; 27.6%)	0.52 (NS)
Admitted through general ward or emergency department, n (%)	730 (of 962; 75.9%)	688 (of 904; 76.1%)	42 (of 58; 72.4%)	0.84 (NS)
Positive blood cultures, patients, n ^{††}	28	25	3 (HRMO <i>E.coli</i> 2, MRSA 1)	
LOS ICU, days, median (range)	1 (1-130)	1 (1 - 88)	5 (1 - 130)	< 0.001
ICU mortality, patients, n (%)	74 (7.7%)	63 (7.0%)	11 (19.0%)	0.0031

^{††} Blood cultures with common skin contaminants (e.g. coagulase-negative Staphylococci, viridans group Streptococci) had to be cultured on two or more separate occasions to be included (n = 23 cultures with positive culture with skin contaminant on one occasion excluded); HRMO = highly resistant microorganism; LOS = length of stay.

Table 4. HRMO species and patient characteristics with HRMO

HRMO, patients, n (% of total patients)	58 (6.0%)
HRMO, total, n*	60
Enterobacteriaceae	50
<i>E.coli</i>	40
<i>Klebsiella</i> spp.	2
Other [†]	8
Non-fermenting gram-negatives	5
<i>Pseudomonas</i> spp.	4
Other [‡]	1
Gram-positives	5
VRE	3
MRSA	2
LOS ICU on first day of positive HRMO culture	
Days, median (range)	1 (1-77)
1 day, n (% of 60)	36 (60%)
2-7 days, n (% of 60)	12 (20%)
8-14 days, n (% of 60)	4 (6.7%)
> 14 days, n (% of 60)	8 (13.3%)
HRMO patients not referred from elsewhere and HRMO within three days (% of 58)	36 (62.1%)
Admitted in preceding 3 months (n,% of 36)	27 (75.0%)
Recent antibiotic exposure (n,% of 36)	12 (33.0%)
Comorbid conditions (n,% of 36)	32 (88.9%)
Cardiovascular (n,% of 36)	11 (30.6%)
Malignancy (n,% of 36)	10 (27.8%)
Organ transplantation (n,% of 36)	7 (19.4%)
Pulmonary (n,% of 36)	6 (16.7%)
Diabetes (n,% of 36)	4 (11.1%)
Chronic hepatitis (HCV, HBV ^{††}) (n,% of 36)	2 (5.6%)
Occupational exposure (pig farmer) (n,% of 36)	2 (5.6%)
No known risk factor for HRMO (n,% of 36)	2 (5.6%)

*2 patients had 2 HRMOs. [†]*E. cloacae* 4; *Citrobacter* spp. 3; *S. marcescens* 1. [‡]*S. paucimobilis* 1; HRMO = highly resistant microorganism; LOS = length of stay; HBV = hepatitis B virus; HCV = hepatitis C virus.

to the ICU; 12 patients (33.3%) had received antibiotics in the months preceding ICU admission. Two patients (of 36, 5.6%) were farmers working with livestock (pigs); both were found to be methicillin-resistant *Staphylococcus aureus* (MRSA) positive. Two (of 36, 5.6%) had no comorbid conditions, no recent hospital admission, no recent antibiotic treatment and no occupational exposure to HRMOs.

Median LOS for all ICU patients was 1 day (range 1-130 days, mean 4 days). LOS in the ICU for HRMO-positive patients was significantly longer (median 5 days, range 1-130, mean 16 days) compared with HRMO-negative patients (median 1 day, range 1-88, mean 3 days) ($p = 0.000$) (table 3).

LOS in the ICU at the time of first positive culture for any HRMO was 1 day in 36 (60%), 2-7 days in 12 (20%), 8-14 days in 4 (7%) and more than 14 days in 8 (13%) (table 4). Patient categories with most HRMO-positive patients were medical (22 out of 208, 10.6%), surgical (14 out of 181, 7.7%) and trauma (6 out of 42, 13.6%). Units with most HRMO-positive patients were the surgical unit (27 out of 218, 11.3%) and the medical unit (13 out of 181, 7.2%).

Of patients admitted to our ICU for more than 14 days, 18 of 57 (31.6%) were found to have any HRMO during ICU stay vs. 31 of 840 (3.7%) patients staying 7 days or less in our ICU ($p < 0.0001$).

APACHE II score for HRMO-positive patients (available in 57 patients) was significantly higher (median 17, mean 19, range 2-52) compared with the APACHE II score for HRMO-negative patients (available in 875 patients) (median 13, mean 13, range, 2-44) ($p = 0.000$).

Overall ICU mortality was 74 (7.7%); mortality was significantly higher in patients with HRMO (11 out of 58,

19.0%) than patients without HRMO (63 out of 904, 7.0%) ($p = 0.0031$) (table 3).

Further, 25 patients had a positive blood culture with a susceptible microorganism and three other patients had a positive blood culture with an HRMO (*E. coli* 2, MRSA 1). In the readmitted (excluded) patient group ($n = 99$), 16 patients (16.2%) had any HRMO (*E. coli* 6, *Klebsiella* spp. 3, other Enterobacteriaceae 3, *Pseudomonas* spp. 1, other non-fermenting Gram-negatives 1, vancomycin-resistant Enterococci 3). This percentage of HRMO-positive patients is significantly higher compared with the percentage of HRMO-positive patients in the study group (6.0%, $p = 0.0002$).

In this study, we could not identify patient characteristics with sufficient specificity and sensitivity to predict HRMO carriage. During the study period, we did not find clusters of identical HRMOs indicating an outbreak.

DISCUSSION

In this single-centre prospective study on the burden of HRMOs in critical care patients in an area where HRMOs are non-endemic,^{8,13} it is an important finding that more than half of HRMO-positive patients were identified from cultures taken on admission. This finding suggests that an important part of antibacterial resistance is imported to the ICU, rather than acquired during the ICU stay. Indeed, hospitalisation on a general ward prior to ICU admission is a recognised risk factor for HRMO acquisition¹⁴ and although the proportion of HRMOs introduced onto the ICU through already colonised or infected patients has been quantified in studies for MRSA,¹⁵ its contribution for all HRMOs has, to the best of our knowledge, not been clearly elucidated as yet in our region. Of all patients admitted through the emergency department or general ward, 36 (out of total 932, 3.7%) had an HRMO within three days of ICU stay. Two (of 36, 5.6%) of these patients had no comorbid conditions, no recent hospital admission, no recent antibiotic treatment and no occupational exposure to HRMOs. Although a minority, this underscores the fact that HRMO is not restricted to the hospital, even in our area of low antibiotic resistance. Indeed, prevalence of antimicrobial resistance in the community, for instance carriage of extended-spectrum beta-lactamases-producing Enterobacteriaceae, is considerable, where contribution of contaminated foods – mainly poultry – and travel, remains to be elucidated.^{9–11}

Only three patients had a proven infection (bacteraemia) with any HRMO, therefore colonisation appears to be far more frequent than a serious infection. That HRMO-positive patients have a higher mortality might in part be explained by the fact that sicker patients are more

often colonised with any HRMO. Indeed, in our study population those with an HRMO had a significantly higher APACHE-II score than those without HRMO (table 2).

In our cohort, Gram-negative bacteria comprised the largest part of all HRMOs. This is in line with the trend towards more Gram-negative antibiotic resistance in European ICUs, with a stabilisation or decrease in Gram-positive antibiotic resistance.^{1,16} Recently, others described cephalosporin and aminoglycoside resistance in a substantial number of critically ill patients colonised with Enterobacteriaceae (15 and 10%, respectively) on ICU admission in a large Dutch multi-centre trial.¹⁷

We could not identify clusters of HRMO. Therefore, the hygiene measures set forth to contain the spread of HRMOs appeared sufficient in this study period.

Our finding that a substantial part of HRMOs are imported into the ICU underscores the imperative need to ensure strict application of hygienic practices, such as hand washing, as well as excellent use of antibiotic stewardship throughout medical care, inside and outside the hospital. Furthermore, this finding highlights the importance of a conscientious surveillance program on the ICU. Along this line, surveillance screening *before* possible ICU admission in specific populations on medical and surgical wards, or in patients with a high risk of community-acquired HRMO, could be worthwhile in order to prevent cross-contamination on these wards and on the ICU. Indeed, in the Netherlands, a surveillance program comparable to our practice on the ICU is carried out on haemato-oncology and dialysis wards as well. Likewise, it is common practice to screen those with occupational exposure to livestock known to have a high carriage rate of MRSA prior to or at hospital admission.¹⁸ Expanding surveillance to other high-risk populations, for instance those admitted to a surgical ward for an extended period, especially when receiving antibiotics, might be beneficial. Some variables have been recognised as risk factors for carriage of HRMOs upon ICU admission, such as prior antimicrobial treatment, prior hospitalisation, and residence in a nursing home.⁵ However, due to sample size, we could not extract patient characteristics from our study population to predict HRMO carriage. It might be helpful to further characterise those patients at risk for HRMO carriage in the context of a larger epidemiological study. This could lead to more differentiated isolation procedures and a more sophisticated choice of antibiotics in case of a proven or suspected infection. Further studies, however, do necessitate a uniform definition of 'highly resistant' enabling meaningful comparisons and data aggregation; currently, a wealth of definitions are being applied in the literature.¹⁹

There are some limitations to our study, the most important being that it is a single-centre study with a relatively small sample size. It is important to note that

patients with an intended stay of two days or less are not included in the surveillance program, unless transferred from another hospital; positive cultures could thus have been missed in these patients. In this group of patients with a short LOS in the ICU, these potential false-negatives would contribute to our finding that a substantial number of HRMO-positive patients are found to be as such in the first days of admission. As our surveillance program is robustly implemented in our daily practice, it is unlikely that patients were missed out of this program for procedural reasons. Some patients underwent surveillance screening despite a short stay of 48 hours or less on the ICU. We did not differentiate between colonisation and infection with any HRMO. Causality between occurrence of an infection with an HRMO and worse outcome in the HRMO group can thus not be proven in this observational study. We did not analyse all known risk factors for HRMO carriage (referral from a nursing home was not recorded); due to sample size, subgroup analyses were not feasible. It could be of benefit to further characterise those at risk of harbouring infection or colonisation with an HRMO, using a uniform international definition of HRMO, with expanding surveillance in high-risk groups outside the ICU, in order to enable maximum precautionary measures and give optimal antibiotic treatment. This combination of surveillance and timely isolation can prevent further spread of HRMOs, our biggest challenge in infection control in critically ill patients in the years ahead.

CONCLUSION

This observational study suggests that HRMOs on the ICU are quite often imported and not only acquired during the stay in the ICU. Gram-negative HRMOs were more abundant than Gram-positive and are of clinical significance even in a non-endemic area. Although most patients with any HRMO had comorbid conditions, were recently admitted to a hospital, had received antibiotics prior to ICU admission, or had occupational exposure to an HRMO, a small minority had no relevant history. Our findings underscore the importance of infection control and optimal surveillance on admission to the ICU.

ACKNOWLEDGEMENTS

We would like to thank Mr. M. Lazonder for building the database and Mr. H. van Assen for providing APACHE II scores of included patients. Furthermore, we wish to thank Ms. K. Glatman for correcting the English text.

DISCLOSURES

The authors declare no conflicts of interest. No funding or financial support was received.

REFERENCES

1. Brusselaers N, Vogelaers D, Blot S. The rising problem of antimicrobial resistance in the intensive care unit. *Ann Intensive Care*. 2011;1:47.
2. Vincent JL, Rello J, Marshall J, et al. International study of the prevalence and outcomes of infection in intensive care units. *JAMA*. 2009;302:2323-9.
3. Kollef MH, Fraser VJ. Antibiotic resistance in the intensive care unit. *Ann Intern Med*. 2001;134:298-314.
4. Bonten MJ. Infection in the intensive care unit: Prevention strategies. *Curr Opin Infect Dis*. 2002;15:401-5.
5. Nseir S, Grailles G, Soury-Lavergne A, Minacori F, Alves I, Durocher A. Accuracy of american thoracic Society/Infectious diseases society of america criteria in predicting infection or colonization with multidrug-resistant bacteria at intensive-care unit admission. *Clin Microbiol Infect*. 2010;16:902-8.
6. de Kraker ME, Davey PG, Grundmann H, BURDEN study group. Mortality and hospital stay associated with resistant *Staphylococcus aureus* and *Escherichia coli* bacteremia: Estimating the burden of antibiotic resistance in Europe. *PLoS Med*. 2011;8:e1001104.
7. Jabes D. The antibiotic R&D pipeline: An update. *Curr Opin Microbiol*. 2011;14:564-9.
8. European Antimicrobial Resistance Surveillance Network (EARS-Net) [Internet]. Stockholm: European Centre for Disease Prevention and Control (ECDC); 2010. Available from: www.ecdc.europa.eu/en/activities/surveillance/EARS-Net/.
9. Leverstein-van Hall MA, Dierikx CM, Cohen Stuart J, et al. Dutch patients, retail chicken meat and poultry share the same ESBL genes, plasmids and strains. *Clin Microbiol Infect*. 2011;17:873-80.
10. Reuland EA, Overdeest IT, Al Naiemi N, et al. High prevalence of ESBL-producing enterobacteriaceae carriage in Dutch community patients with gastrointestinal complaints. *Clin Microbiol Infect*. 2013;19:542-9.
11. Huijbers PM, de Kraker M, Graat EA, et al. Prevalence of extended-spectrum beta-lactamase-producing enterobacteriaceae in humans living in municipalities with high and low broiler density. *Clin Microbiol Infect*. 2013;19:E256-9.
12. Kluytmans-Vandenbergh MF, Kluytmans JA, Voss A. Dutch guideline for preventing nosocomial transmission of highly resistant microorganisms (HRMO). *Infection* 2005;33:309-13.
13. de Kraker M, van de Sande-Bruinsma N. Trends in antimicrobial resistance in Europe: Update of EARSS results. *Euro Surveill*. 2007;12:E0703153.
14. Vogelaers D, De Bels D, Foret F, et al. Patterns of antimicrobial therapy in severe nosocomial infections: Empiric choices, proportion of appropriate therapy, and adaptation rates--a multicentre, observational survey in critically ill patients. *Int J Antimicrob Agents*. 2010;35:375-81.
15. Merrer J, Santoli F, Appere de Vecchi C, Tran B, De Jonghe B, Outin H. "Colonization pressure" and risk of acquisition of methicillin-resistant staphylococcus aureus in a medical intensive care unit. *Infect Control Hosp Epidemiol*. 2000;21:718-23.
16. van Duijn PJ, Dautzenberg MJ, Oostdijk EA. Recent trends in antibiotic resistance in European ICUs. *Curr Opin Crit Care*. 2011;17:658-65.
17. Oostdijk EA, de Smet AM, Kesecioglu J, Bonten MJ, on behalf of the Dutch SOD-SDD Trialists Group. Decontamination of cephalosporin-resistant enterobacteriaceae during selective digestive tract decontamination in intensive care units. *J Antimicrob Chemother*. 2012;67:2250-3.
18. Van Cleef BA, Broens EM, Voss A, et al. High prevalence of nasal MRSA carriage in slaughterhouse workers in contact with live pigs in the Netherlands. *Epidemiol Infect*. 2010;138:756-63.
19. Souli M, Galani I, Giamarellou H. Emergence of extensively drug-resistant and pandrug-resistant gram-negative bacilli in Europe. *Euro Surveill*. 2008;13.