

Familial LCAT deficiency: from renal replacement to enzyme replacement

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ABSTRACT

Familial LCAT deficiency (FLD) is a recessive lipid disorder ultimately leading to end-stage renal disease (ESRD). We present two brothers with considerable variation in the age at which they developed ESRD. Kidney biopsies revealed both tubular and glomerular pathology. To date, no causal therapy is available, yet enzyme replacement therapy is in development.

KEYWORDS

Familial LCAT deficiency, FLD, LCAT, renal failure

What was known on this topic?

Familial LCAT deficiency (FLD) leads to end-stage renal disease (ESRD). No causal therapy is available to date.

What does this add?

We show considerable variation in the age at which ESRD occurs in FLD, demonstrate that FLD causes tubular pathology besides glomerular pathology, and discuss the development of enzyme-replacement therapy.

INTRODUCTION

Patients with familial lecithin-cholesterol acyltransferase (LCAT) deficiency (FLD), a rare autosomal recessive disorder, are characterised by progressive corneal opacification, glomerulopathy, mild haemolytic anaemia and very low plasma levels of high-density lipoprotein cholesterol (HDL-c).¹ The molecular defect underlying FLD is homozygosity or compound heterozygosity for mutations in the gene encoding LCAT. LCAT is a pivotal enzyme in cholesterol metabolism, by virtue of its ability to esterify cholesterol molecules in HDL and low density-lipoprotein (LDL) particles, anchoring them in the lipophilic cores of these lipoproteins.^{2,3}

With no causal therapy currently available, FLD patients often develop end-stage renal disease in the fourth decade of life.⁴ Here, we describe FLD in two brothers of Turkish descent. One recently received a kidney transplant, the other developed severe renal insufficiency. The described cases illustrate the need for enzyme-replacement therapy, currently in preclinical development.⁵

CASE REPORT

Patient 1 was referred at the age of 25 because of generalised oedema. The patient had bilateral corneal opacification. Upon physical examination signs of lung oedema were noted. Blood pressure was 140/95 mmHg. Clinical characteristics are depicted in *figure 1*. Blood sample analysis revealed increased glomerular filtration and hypoalbuminaemia, complete HDL-C deficiency and low apolipoprotein A-I. In addition, a mild microcytic anaemia was shown. Proteinuria was noted in the absence of leucocyturia or erythrocyturia. Upon ultrasonography, no structural renal abnormalities were found.

Quinapril, furosemide and candesartan were started, resulting in resolution of the oedema and a decrease in proteinuria and renal clearance (12.4 g/l to 9.4 g/l and from 165 ml/min to 130 ml/min). Notwithstanding, the

Figure 1. Pedigree of FLD patients

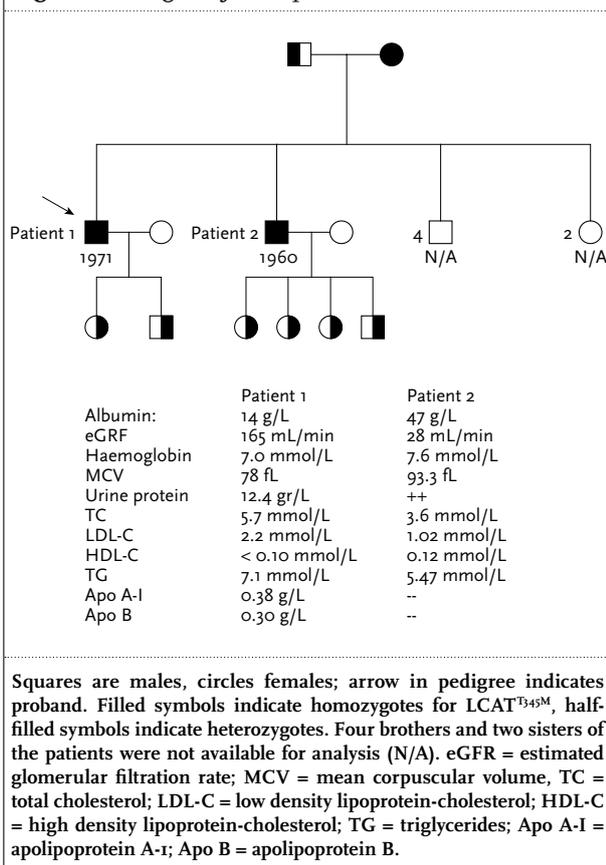
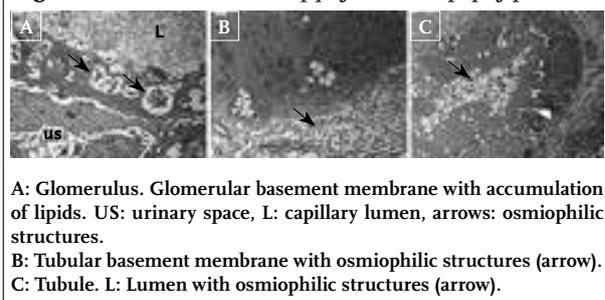


Figure 2. Electron microscopy of renal biopsy of patient 1



DISCUSSION

Here we report familial $LCAT$ deficiency (FLD) in two brothers of Turkish descent. We show that the age at which end-stage renal disease develops is variable and we also demonstrate that renal pathology in FLD is not only characterised by glomerular changes, but also by profound tubular changes.

To date, 88 mutations in $LCAT$ have been described.⁸ Severe mutations lead to complete inactivation of the $LCAT$ enzyme, and consequentially to FLD. The $T345M$ mutation identified in our two patients was originally described in a Sardinian patient.⁷ The threonine residue at position 345 is conserved up to *C. elegans*, but it is currently unknown why this specific residue is crucial for normal enzyme function.

Patients with FLD suffer from progressive proteinuria and renal dysfunction, on average leading to ESRD in the fourth decade of life.¹ Patient 1 indeed developed ESRD at the age of 37, but Patient 2, his full brother, was 50 years old when severe renal insufficiency was noted during a work-up for renal transplantation. The reason for the variable course of renal disease progression in our two FLD patients is unknown and remarkable given the fact that the two brothers had a comparable lifestyle; neither had any comorbidity influencing renal function such as diabetes mellitus, hypertension or cardiovascular disease.

The exact pathogenesis of renal disease in FLD is unknown. Due to the absence of functional $LCAT$, excess unesterified cholesterol and phospholipid are present in these patients, leading to the formation of lipoprotein X, a protein-free aberrant particle containing FC and PL, associated with glomerular endothelial damage.⁹ The composition of typical vacuoles in the glomerular basement of FLD patients has not been fully characterised to date.⁹⁻¹⁰ The presence of foam cells in the mesangium of our patient, however, suggests that excess lipid is important in the pathogenesis of the renal damage observed in our FLD patient.

patient developed ESRD at the age of 37. His brother – Patient 2, 50 years old – was evaluated as a kidney donor, but upon assessment he was shown to suffer from corneal opacification, HDL-C deficiency and severe proteinuria as well.

Electron microscopy of a renal biopsy of Patient 1 showed accumulation of electron-lucent vacuoles with or without osmiophilic particle cores in the glomerular basement membrane, Bowman's capsule, the tubular basement membrane and the lumen of tubules. Segmental foot process effacement, foam cell accumulation in the mesangium and a distorted architecture of the glomerular and tubular basement membrane were also noted (figure 2).

The patients' parents were first cousins (figure 1), and the presence of FLD as underlying disease was anticipated. Plasma $LCAT$ activity, measured as the ability of plasma $LCAT$ to esterify free cholesterol in proteoliposomes,⁶ was severely reduced in both patients: 0.75 nmol cholesterol ester/h/ml (normal: 25 nmol/h/ml). Upon DNA analysis,³ both patients were found to be homozygous carriers of an $ACG \rightarrow ATG$ mutation in exon 6, resulting in the p.T345M substitution in $LCAT$.

To date no causal treatment for FLD is available. As a consequence, symptomatic treatment encompassing lipid lowering and antihypertensive therapy is commonly started in FLD patients. This combination has been shown to result in decreased proteinuria and stabilisation of pathological sequelae in FLD patients.¹¹ Beneficial effects of corticosteroid administration have also been described.¹² FLD patients who develop ESRD require haemodialysis or kidney transplantation. Despite evidence of early histomorphological changes consistent with FLD in renal grafts after transplantation, acceptable long-term results have been reported.¹³

Both LCAT gene replacement and enzyme replacement are under development. In a model described by Kuroda and co workers,¹⁴ autologous adipocytes are transfected with human LCAT via a retroviral vector. LCAT secreted by these cells is able to restore enzyme activity in plasma of FLD patients. Recombinant human LCAT (ACP-501)¹⁵ showed excellent results in LCAT knockout mice, rapidly restoring LCAT activity, cholesterol efflux and lipid profiles. rhLCAT is currently being evaluated in FLD patients in a phase I trial.⁵

CONCLUSION

FLD is a recessive lipid disorder ultimately resulting in ESRD. The clinical course of the disease is variable, even in related FLD patients. We demonstrate that apart from glomerulopathy, FLD is also characterised by tubular pathology. Finally, our patients stress the need for LCAT enzyme replacement therapy, which is currently under development in a phase I clinical trial.⁵

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REFERENCES

1. Holleboom AG, Kuivenhoven JA, van Olden CC, et al. Proteinuria in early childhood due to familial LCAT deficiency caused by loss of a disulfide bond in lecithin:cholesterol acyl transferase. *Atherosclerosis*. 2011;216:161-5.
2. Lewis G, Rader D. New insights into the regulation of HDL metabolism and reverse cholesterol transport. *Circ Res*. 2005;96:1221-32.
3. Holleboom AG, Kuivenhoven JA, Peelman F, et al. High prevalence of mutations in LCAT in patients with low HDL cholesterol levels in the Netherlands: Identification and characterization of eight novel mutations. *Hum Mutat*. 2011;32:1-9.
4. Santamarina-Fojo S, Lambert G, Hoeg JM, Brewer HB Jr. Lecithin cholesterol acyltransferase: role in lipoprotein metabolism, reverse cholesterol transport and atherosclerosis. *Curr Opin Lipidol*. 2000;11:267-75.
5. <http://www.alphacorepharma.com/wp-content/uploads/2012/02/ACP-ABL-press-release-final.pdf>. Internet source.
6. Fröhlich J, McLeod R, Pritchard PH, Fesmire J, McConathy W. Plasma lipoprotein abnormalities in heterozygotes for familial lecithin:cholesterol acyltransferase deficiency. *Metabolism*. 1988;37:3-8.
7. Funke H, von Eckardstein A, Pritchard PH. Genetic and phenotypic heterogeneity in familial lecithin: cholesterol acyltransferase (LCAT) deficiency. Six newly identified alleles further contribute to the structural heterogeneity in this disease. *J Clin Invest*. 1993;91:677-83.
8. Human Gene Mutation Database. Human Gene Mutation Database. 2010. Ref Type: Internet Communication.
9. Imbasciati E, Paties C, Scarpioni L, Mihatsch MJ. Renal lesions in familial lecithin-cholesterol acyltransferase deficiency: ultrastructural examination of glomerular changes. *Am J Nephrol*. 1986;6:66-70.
10. Jonas A. Regulation of lecithin cholesterol acyltransferase activity. *Prog Lipid Res*. 1998;37:209-4.
11. Aranda P, Valdivielso P, Pisciotto L, et al. Therapeutic management of a new case of LCAT deficiency with a multifactorial long-term approach based on high doses of angiotensin II receptor blockers (ARBs). *Clin Nephrol*. 2008;69:213-8.
12. Miarka P, Idzior-Waluś B, Kuźniewski M, Waluś-Miarka M, Klupa T, Sutowicz W. Corticoid treatment of kidney disease in a patient with familial lecithin-cholesterol acyltransferase deficiency. *Clin Exp Nephrol*. 2011;15:424-9.
13. Panescu V, Grignon Y, Hestin D, et al. Recurrence of lecithin cholesterol acyltransferase deficiency after kidney transplantation. *Nephrol Dial Transplant*. 1997;12:2430-2.
14. Kuroda M, Aoyagi Y, Asada S, et al. Ceiling Culture-Derived Proliferative Adipocytes are A Possible Delivery Vehicle for Enzyme Replacement Therapy in Lecithin: Cholesterol Acyltransferase Deficiency. *Open Gene Ther J*. 2011;4:1-10.
15. Rousset X, Vaisman B, Auerbach B, et al. Effect of recombinant human lecithin cholesterol acyltransferase infusion on lipoprotein metabolism in mice. *J Pharmacol Exp Ther*. 2010;335:140-8.