Bleomycin and scuba diving: to dive or not to dive?

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ABSTRACT

Bleomycin is to treat patients with testicular cancer and lymphoma. Bleomycin can bind to DNA and chelate iron. The resulting complex can form an intermediate capable of interacting with oxygen to produce reactive oxygen species, particularly superoxide. Administrating high-inspired oxygen concentrations (e.g. during anaesthesia or acute illness) has been reported to exacerbate pulmonary injury. The duration of risk after bleomycin chemotherapy is unknown. Here we discuss our advice to a young male patient, who was successfully treated with bleomycin for testicular cancer, concerning the safety to return to scuba diving. Since scuba divers are exposed to high partial oxygen pressures (depending on the depth of the dive) we discouraged this patient from resuming scuba diving.

INTRODUCTION

Bleomycin is given in the standard regimen for treating testicular cancer. The incidence of fatal pulmonary toxicity in this low-risk population of young male patients is approximately 2 to 3%. Patients treated with bleomycin are sensitive to oxygen-mediated lung injury. Here we discuss a question patients frequently ask concerning the safety of returning to scuba diving following bleomycin therapy.

CASE REPORT

A 35-year-old man was admitted to our hospital because of a painless enlargement of the left testis, without further symptoms. Ultrasonography demonstrated a solid mass in the left testis. Laboratory examination showed an elevated serum chorionic gonadotropin (β-hCG) of 1100 IU/l (normal: <2) and a serum lactate dehydrogenase of 520 IU/l (normal: 160-320); serum alfa-foetoproteine was normal. Chest X-ray was normal. CT scan of the abdomen showed retroperitoneal lymphadenopathy (maximal diameter 6 cm). The patient proceeded to inguinal orchidectomy with removal of the affected testis. Histopathologically the removed testis consisted of choriocarcinoma. Since this patient had a stage IIC carcinoma with a good prognosis, he was treated with three courses of BEP (bleomycin, etoposide and cisplatin). No pulmonary toxicity was observed. Subsequent radiographic evaluation showed no residual disease and tumour markers normalised. At this time our patient, who used to be an active scuba diver, asked whether it was safe to return to scuba diving after having undergone chemotherapy containing bleomycin. We will discuss our considerations based on a literature search.

BLEOMYCIN

The bleomycins are a family of cytotoxic glypeptide antibiotics isolated from Streptomyces verticullis, with a molecular weight of approximately 1500 D. All contain a unique structural component, bleomycinic acid, and differ only in their terminal alkylamine group. Bleomycin A2, the predominant peptide, and a series of analogues are prepared by total chemical synthesis.¹ The primary biochemical action of the A2 peptide is to produce single- and double-strand breaks in DNA. This breakage is reflected in the chromo-

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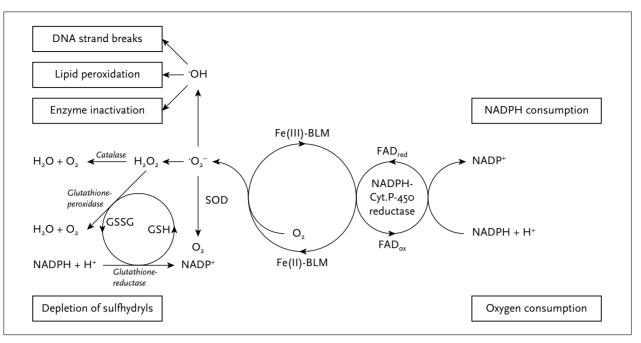


Figure 1

Redox cycling of the iron bleomycin complex with subsequent 'oxidative stress' caused by the formation of reactive oxygen species (superoxide radical O_2 -, hydroxyl radical OH, hydrogen peroxide H_2O_2), potentially toxic reactions and enzymatic detoxification mechanisms

SOD = superoxide dismutase, GSSG = oxidised glutathione, GSH = glutathione, NADPH = nicotinamide adenine dinucleotide phosphate (reduced form), NADP+ = nicotinamide adenine dinucleotide phosphate (oxidised form), FADred = flavin adenine dinucleotide (reduced form), FADox = flavin adenine dinucleotide (oxidised form), BLM = bleomycin.

somal gaps, deletions and fragments seen in cytogenetic studies of whole cells. The mechanism of DNA breakage has been clarified by investigation of the action of bleomycin on both viral and mammalian DNA and results from the production of free radicals by an Fe(II)-bleomycin complex intercalated between opposing strands of DNA (see figure 1). The initial step in this reaction sequence seems to be the production of an activated bleomycin-Fe(II)-O2 complex. The activated complex then binds to DNA. At saturating concentrations of bleomycin, one molecule of drug is bound per four to five base pairs of DNA.2 The second step in the action of bleomycin is the induction of DNA breaks, mediated by free radicals produced by the activated bleomycin-Fe(II) complex. The importance of Fe(II) is indicated by the observation that iron-chelating agents inhibit the DNA scission reaction.3 The enzyme-like bleomycin-Fe(II) complex induces the reduction of molecular oxygen to superoxide and hydroxyl radicals.⁴ In this process, Fe(II) undergoes oxidation to Fe(III). The hypothesis that oxygen radicals participate in the DNA cleavage mediated by bleomycin is based on several observations. First, strand breakage requires the presence of O₂ and ceases in an anaerobic environment.^{5,6} Second, the oxidation of bleomycin-Fe(II) requires oxygen consumption.3

Finally free-radical scavengers and superoxide dismutase (which inactivate O₂ radicals) inhibit DNA strand breakage in *vitro*⁷ and pulmonary toxicity *in vivo*.⁸

BLEOMYCIN AND PULMONARY TOXICITY

Intracellular bleomycin is inactivated by an aminohydrolase that is found in both normal and malignant cells.⁹ The enzyme cleaves the carboxamide amine from the β -aminoalaninamide, yielding a weakly cytotoxic deamidobleomycin. Interestingly, this enzyme is present in relatively low concentrations in lung and skin, the two normal tissues most susceptible to bleomycin damage.¹⁰ Several distinct pulmonary syndromes have been associated with the use of bleomycin, such as bronchiolitis obliterans with organising pneumonia (BOOP), eosinophilic hypersensitivity and, most commonly, interstitial pneumonitis.¹¹ In its later stages interstitial pneumonitis can be complicated by progressive interstitial fibrosis, hypoxia and death. Pulmonary toxicity, usually manifesting with cough, dyspnoea and bibasilar pulmonary infiltrates on chest X-ray film, occurs in 3 to

5% of patients receiving a total dose of less than 450 units of bleomycin, increasing significantly to a 10% incidence in those treated with greater cumulative doses.¹² Although the risk of lung toxicity increases with cumulative doses greater than 450 units, severe pulmonary sequelae have been observed at total doses below 100 units. In the standard regimen for treating testicular cancer, bleomycin is given in doses of 30 units weekly for 9 to 12 doses, and the incidence of fatal pulmonary toxicity in this low-risk population of young male patients is about 2 to 3%.13,14 Pulmonary function tests, particularly the carbon monoxide diffusing capacity, are of possible value in predicting a high risk of pulmonary toxicity. However, most patients treated with bleomycin show a progressive (10 to 15%) deterioration in diffusion capacity with increasing total dose and a more marked increase in changes above 240 units total dose. It is not clear whether the diffusion capacity test can be used to predict which patients will subsequently develop clinically significant pulmonary toxicity.¹⁵ In advanced stages in the evolution of bleomycin pulmonary toxicity, the diffusion capacity as well as arterial O₂ saturation and total lung capacity become markedly abnormal. Besides the total dose of bleomycin given, various other factors have shown to increase the pulmonary toxicity of bleomycin: prior radiation of the lung parenchyma,^{16,17} administration of high fractional-inspired oxygen concentration,¹⁸⁻²² the age of the patient²³ and renal insufficiency (bleomycin is cleared by the kidneys).²⁴ The sensitivity of bleomycin-treated patients to high concentrations of inspired O₂ is intriguing in view of the molecular action of bleomycin, which is dependent on, and mediated by, the formation of oxygen-derived free radicals. Goldliner et al. observed five testicular tumour patients treated with 135 to 595 units of bleomycin 7 to 12 months earlier who underwent retroperitoneal lymph node dissection or resection of pulmonary metastases while receiving an intraoperative fractional concentration of inspired oxygen (FIO₂) ranging from 0.35 to 0.42.18 All five developed respiratory failure postoperatively and died.¹⁸ A reduction in inspired O₂ to an FIO₂ between 0.22 and 0.25, and a decrease in fluids administered during surgery, prevented mortality in subsequent patients.¹⁸ It has been shown that a greater degree of experimental lung injury with oxygen was found at 8 versus 21 days following intratracheal bleomycin.25 Nevertheless, in the abovementioned report by Goldiner et al. the mean time between bleomycin administration and surgery was 9.6 months.¹⁸ Thus, the period of time when oxygen administration appears to be safe following bleomycin has not been established. Therefore, current safeguards for anaesthesia of bleomycintreated patients (both with a history of bleomycin toxicity and even those with previous drug exposure without clinical toxicity) include the use of the minimal tolerated concentration of inspired oxygen and modest fluid replacement to prevent pulmonary oedema.²⁶

SCUBA DIVING

Very few scuba divers, and even fewer sport scuba divers, use oxygen in their tanks. The vast majority of sport divers use compressed air (21% oxygen). The partial inspiratory oxygen pressure (PIO₂) is a function of the fractional concentration of inspired oxygen (FIO₂), the barometric pressure (PB), and the partial pressure of water vapour (PH₂O) in humidified gas; that is PIO₂ = FIO₂ (PB – PH₂O).²⁷ So the partial pressure of oxygen in the inspired (compressed) air is a direct function of the depth of the dive. For every 9.9 m depth of a seawater dive, the ambient barometric pressure to which the diver is exposed increases by I atm. At a dive depth of 19.8 m of seawater (3 atm total pressure), the partial pressure of inspired oxygen in a scuba diver breathing compressed air is 0.63 atm, equivalent to breathing 63% oxygen on the surface. At a dive depth of 29.7 m of seawater, not an unusual depth for many sport divers, the partial pressure of oxygen is 0.84 atm, equivalent to breathing 84% oxygen on the surface.

ADVICE REGARDING PREVIOUS TREATMENT WITH BLEOMYCIN AND SCUBA DIVING

Several clinical and animal studies strongly support the relationship between bleomycin toxicity and oxygen therapy. However, there are no data on what time interval is safe between the last dose of bleomycin and oxygen therapy. Bleomycin is especially successful in the BEP (bleomycin, etoposide, cisplatin) regimen against testicular cancer. These young men frequently ask whether it is safe to resume scuba diving. However, the partial pressure of inspired air is dependent on the barometric pressure, which is a direct function of the depth of the dive. Therefore, these patients should be advised that theoretically there is a risk of developing pulmonary damage due to exposure to a higher partial pressure of inspired oxygen while scuba diving. However, published data regarding the safety of exposure to high concentration oxygen during recreational activities such as scuba diving are limited and patients should be counselled that safety cannot be assured during these activities.²⁸ Since the period of time when oxygen administration appears to be safe following bleomycin has not been established (see above) we think scuba diving should be discouraged, even several years after bleomycin treatment.

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REFERENCES

- Chabner BA, Collins JM (editors). Cancer Chemotherapy. In: Principles & Practice. Philadelphia: Lippincott Company, 1990.
- Umezawa H, Takita T, Sugiura Y, et al. DNA-bleomycin interaction: nucleotide sequence-specific binding and cleavage of DNA by bleomycin. J Antibiot 1978;31:1316-20.
- Sausville EA, Peisach J, Horwitz SB. Effects of chelating agents and metal ions on the degradation of DNA by bleomycin. Biochemistry 1987;17:2740-6.
- Caspary WJ, Niziak C, Lanzo DA, et al. Bleomycin A2: a ferrous oxidase. Mol Pharmacol 1979;16:256-60.
- Onishi T, Iwata H, Takagi Y. Effects of reducing or oxidizing agents on the action of bleomycin. J Biochem 1975;77:745-52.
- Lin SY, Grollman AP. Interactions of a fragment of bleomycin with deoxyribonucleotides: nuclear magnetic resonance studies. Biochemistry 1981;20:7589-98.
- Lown JW, Sim S. The mechanism of the bleomycin-induced cleavage of DNA. Biochem Biophys Res Commun 1977;77:1150-7.
- Pepin JM, Langner RO. Effects of dimethyl sulfoxide (DMSO) on bleomycin induced cleavage of DNA. Biochem Pharmacol 1985;34:2386-9.
- Muller WEG, Zahn RK. Bleomycin: mode of action on DNA. In: Fundamental and clinical studies of bleomycin. Carter SK, Ichikawa T, Mathe G, et al (editors). Baltimore: University Park Press, 1976;51-62.
- Umezawa H, Hori S, Sawa T, et al. A bleomycin-inactivating enzyme in mouse liver. J Antibiot 1974;27:419-24.
- 11. Sleijfer S. Bleomycin-induced pneumonitis. Chest 2001;120:617-24.
- Blum RH, Carter SK, Agre K. A clinical review of bleomycin a new antineoplastic agent. Cancer 1973;31:903-14.
- Williams SD, Birch R, Einhorn LA, et al. Treatment of disseminated germ cell tumors with cisplatin, bleomycin, and either vinblastine or etoposide. N Engl J Med 1987;316:1435-40.
- Simpson AB, Paul J, Graham J, et al. Fatal bleomycin pulmonary toxicity in the west of Scotland 1991-95; a review of patients with germ cell tumours. Br J Cancer 1998;78:1061-6.

- Comis RL, Kuppinger MS, Ginsberg SJ, et al. Role of single breath carbon monoxide-diffusing capacity in monitoring the pulmonary effects of bleomycin in germ-free tumor patients. Cancer Res 1979;39:5076-80.
- Perez-Guerra F, Harkleroad LE, Walsh RE, Constanzi JJ. Acute bleomycin lung. Am Rev Respir Dis 1972;106:909-13.
- Sammuels ML, Johnson DE, Holoye PY, Lazotti VJ. Large-dose bleomycin therapy and pulmonary toxicity: a possible role of prior radiotherapy. JAMA 1976;235:1117-20.
- Goldiner PL, Carlon GC, Critkovic E, et al. Factors influencing post-operative morbidity and mortality in patients treated with bleomycin. BMJ 1978;1:1664-7.
- 19. Gilson AJ, Sahn SA. Reactivation of bleomycin lung toxicity following oxygen administration. Chest 1985;88:304-6.
- 20. Tryka AF, Skornik WA, Godlesti JJ, Brain JD. Potentation of bleomycin-induced lung injury by exposure to 70% oxygen. Am Rev Respir Dis 1982;126:1074-9.
- 21. Ingrassia TS, Ryu JH, Trastek VF, Rosenow EC. Oxygen-exacerbated bleomycin pulmonary toxicity. Mayo Clin Proc 1991;66:173-8.
- 22. Maher J, Daly PA. Severe bleomycin lung toxicity: reversal with high dose corticosteroids. Thorax 1993;48:92-4.
- 23. Yagoda A, Mukherji B, Young C, et al. Bleomycin: an antitumor antibiotic; clinical experience in 274 patients. Ann Intern Med 1972;77:861-70.
- 24. Kawai K, Ninotsu S, Tomobe M, Akaza H. Serum creatinine level during chemotherapy for testicular cancer as a possible predictor of bleomycininduced pulmonary toxicity. Jpn J Clin Oncol 1998;28:546-52.
- 25. Rinaldo J, Goldstein RH, Snider GL. Modification of oxygen toxicity after lung injury by bleomycin in hamsters. Am Rev Resp Dis 1982;126:1030-3.
- 26. Allen SC, Riddel GS, Butchart EG. Bleomycin therapy and anaesthesia. The possible hazards of oxygen administration to patients after treatment with bloemycin. Anaesthesia 1981;36:60-3.
- 27. Marino PL. The ICU book. Baltimore: Lippincott Williams & Wilkins, 1998:339-54.
- 28. Zanetti CL. Scuba diving and bleomycin therapy. JAMA 1990;22:2869.

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