

Clinical significance of soluble interleukin-2 receptor measurement in immune-mediated diseases

W.A. Dik^{1,2,*}, M. Heron³

Departments of ¹Immunology, Laboratory Medical Immunology, ²Internal Medicine, Division Clinical Immunology, Erasmus Medical Center, University Medical Center Rotterdam, the Netherlands;

³Laboratory of Medical Microbiology and Immunology, Diaconessenhuis, Utrecht, the Netherlands.

*Corresponding author: w.dik@erasmusmc.nl

ABSTRACT

A soluble form of the interleukin-2 receptor (sIL-2R) is secreted upon T-cell activation. Increased blood levels of sIL-2R occur in a variety of immunological diseases. Although the biological function of sIL-2R is incompletely understood, both in health and disease, sIL-2R serum measurements are commonly conducted in clinical practice as it may help to facilitate diagnosis of specific immune-mediated diseases, such as haemophagocytic lymphohistiocytosis and sarcoidosis. In these, and in other immune-diseases, sIL-2R levels may be used as a biomarker to monitor/predict disease activity and treatment response. In this review, we will give a brief overview of the biology of the IL-2/IL-2R system and will subsequently discuss the clinical utility of sIL-2R measurement, especially in the context of haemophagocytic lymphohistiocytosis, sarcoidosis, rheumatoid arthritis, systemic lupus erythematosus, juvenile idiopathic arthritis, adult-onset Still's disease, ANCA-associated vasculitis, and IgG4-related disease.

KEYWORDS

Haemophagocytic lymphohistiocytosis, immune-mediated diseases, sarcoidosis, soluble IL-2 receptor, T-cell activation

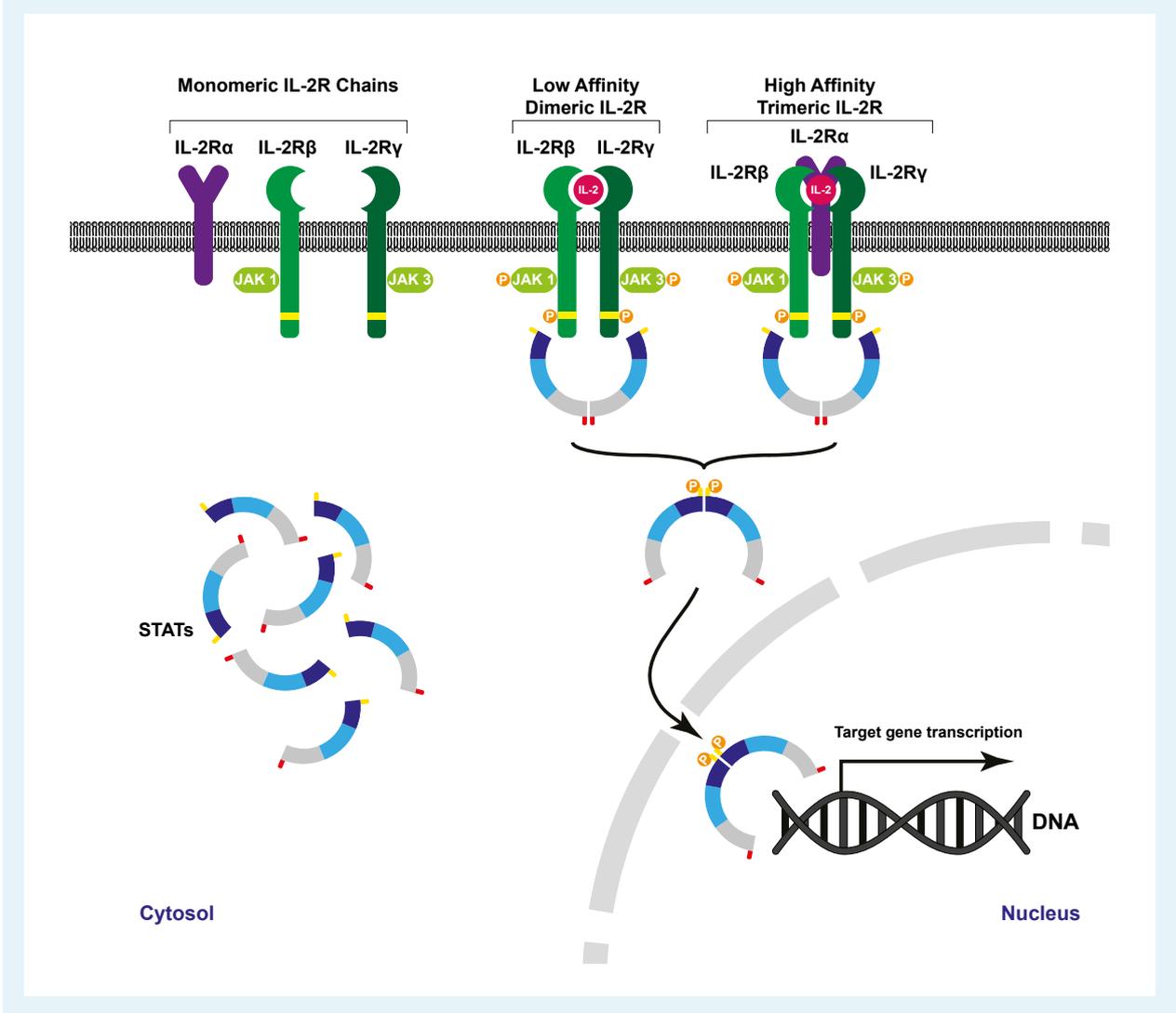
INTRODUCTION

Interleukin 2 (IL-2) represents one of most significant cytokines in the immune system as it is involved in the regulation of protective immunity, as well as maintaining immune tolerance mediated by CD4⁺ regulatory

T lymphocytes (Treg).^{1,3} IL-2 acts on cells that express either the trimeric high-affinity IL-2 receptor (IL-2R) or dimeric low-affinity IL-2R (figure 1). The dimeric low-affinity IL-2R consists of the IL-2R β chain (also known as CD122) and the cytokine receptor common γ -chain (γ_c , also known as CD132). The low-affinity dimeric IL-2R is hardly expressed by naive CD4⁺ T lymphocytes, expressed at low levels by naive CD8⁺ T lymphocytes and memory CD4⁺ T lymphocytes, and at high levels on memory CD8⁺ T lymphocytes and NK cells. Cells that express high levels of low-affinity IL-2R are susceptible to activation by IL-2 in vitro, yet this requires stimulation with (non)-physiological IL-2 concentrations.^{1,4} The third chain of the trimeric high-affinity IL-2R is IL-2R α (also known as CD25 or TAC antigen). IL-2R α does not actively participate in receptor signalling but rather, enhances the receptors affinity for IL-2. Tregs are characterised by strong constitutive expression of IL-2R α which enables these cells to constantly express the high-affinity trimeric IL-2R (IL-2R $\alpha\beta\gamma$) and thereby use the low physiological level of IL-2 as is present in vivo.¹ The high-affinity IL-2R is transiently expressed at high levels by activated CD4⁺ and CD8⁺ T lymphocytes. First, following signalling induced by T-cell receptor (TCR) activation and co-stimulatory molecules, IL-2R α is induced to moderate expression levels which is subsequently further enhanced in a positive feedback loop through IL-2/IL-2R signaling.^{1,3,5}

Rubin and colleagues were the first to demonstrate that after in vitro activation, T lymphocytes not only enhanced cellular IL-2R expression but also released soluble IL-2R(α),⁶ and similar to cellular IL-2R expression, the release of soluble IL-2R required de novo protein synthesis rather than cellular proliferation.⁶ Other studies demonstrated a significant correlation between surface membrane IL-2R expression on activated CD4⁺ and

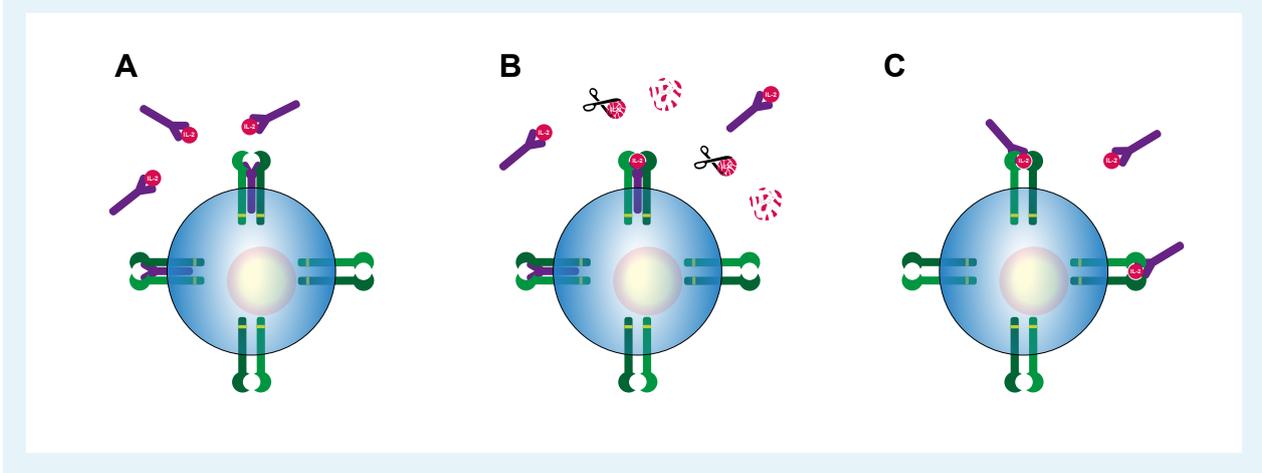
Figure 1. Schematic representation of the IL-2 receptor chains (IL-2R α , IL-2R β , and IL-2R γ), IL-2 binding to the low affinity dimeric IL-2 receptor (comprised of IL-2R β and IL-2R γ), and the high affinity trimeric IL-2 receptor (comprised of IL-2R α , IL-2R β , and IL-2R γ). Binding of IL-2 to the dimeric/trimeric receptor causes a receptor conformational change, activation of receptor-associated janus kinase (JAK) molecules, and creates a docking site for signal transducer and activator of transcription (STAT) molecules. STAT molecules, predominantly STAT5 but also STAT1 and STAT3, are subsequently recruited, phosphorylated and dimerise. Dimeric STAT molecules then translocate to the nucleus where they bind specific DNA sequences and regulate the transcription of many IL-2 target genes.



CD8+ T lymphocytes and the amount of secreted sIL-2R following in vitro activation.⁷ In addition, CD4+CD25+ Tregs were found to secrete sIL-2R upon in vitro activation with certain stimuli.⁸ Soluble IL-2R most likely originates from enzymatic cleavage and release from the cell surface membrane. Several enzymes, including neutrophil-derived elastase, matrix metalloproteinases-9, and the house dust mite protease Der p 1, have been proposed for cleavage of membrane IL-2R α .⁹⁻¹¹ However, although the exact nature of the involved proteolytic factor(s) remains largely unknown, it is most likely endogenous to the

sIL-2R-producing cell as suggested by occurrence of sIL-2R production under serum-free cell culture conditions.^{6,12} Since its initial in vitro description in 1985, elevated sIL-2R blood levels have been found in different pathological conditions, including autoimmune diseases, infectious diseases, transplant rejection, and malignancies.¹³⁻¹⁶ Associations between genetic variants in *IL2RA* (the gene encoding IL-2R α) and autoimmune diseases have been described. However, correlating this directly to serum sIL-2R levels may be difficult given that sIL-2R is produced through membrane cleavage which, in turn,

Figure 2. Proposed mechanisms of action of sIL-2R. A) sIL-2R binds IL-2, thereby prohibiting IL-2 from binding to either to the low affinity dimeric IL-2 receptor or high affinity trimeric IL-2 receptor. B) sIL-2R binds IL-2, thereby protecting IL-2 from enzymatic degradation and prolonging IL-2 half-life. C) sIL-2R binds IL-2, thereby increasing the affinity of IL-2 for the low affinity dimeric IL-2 receptor. IL-2Ra: purple, IL-2R β : light green, IL-2R γ : dark green, sIL-2R: purple.



depends on several different processes which may be influenced by disease activity.¹⁷⁻²² Currently, sIL-2R is generally regarded as a marker of T-lymphocyte activation; however, the cellular source of sIL-2R may not be restricted to T lymphocytes as other types of activated immune cells, including monocytes, dendritic cells, and B lymphocytes may release sIL-2R as well.^{6,16,23-27}

Despite the recognised association between immune activation and increased sIL-2R release under pathological conditions, the biological actions of sIL-2R are still far from understood. Several mechanisms of action, ranging from immune-inhibitory to immuno-stimulatory effects, have been proposed (figure 2). Soluble IL-2R binds IL-2 efficiently, and based on in vitro experiments, it has been proposed that sIL-2R may limit activation and proliferation of T lymphocytes by sequestration of available IL-2.^{1,27-30} However, conflicting data have been reported.⁸ Alternatively, sIL-2R complexed with IL-2 prolongs IL-2 half-life which may enhance the immune-stimulatory properties of IL-2, even by activation of low-affinity dimeric IL-2R.^{31,32} It has been proposed that IL-2 can be presented to CD4⁺ T lymphocytes through sIL-2R, which then induces differentiation into Tregs (rather than differentiation into T-helper (Th)₁ or Th₁₇ lymphocytes) that subsequently can suppress immune activity.³³ On the other hand, there are reports to support observations that sIL-2R may promote (auto)immune processes in association with enhanced Th₁₇ generation, which involves sequestration of the IL-2 that normally inhibits early Th₁₇ differentiation.^{21,34} Although the exact mechanism(s) of action of sIL-2R, as well as their in vivo occurrence and final biological effects,

remains to be determined, the data available so far do support a role for sIL-2R in regulating IL-2-dependent cell function.

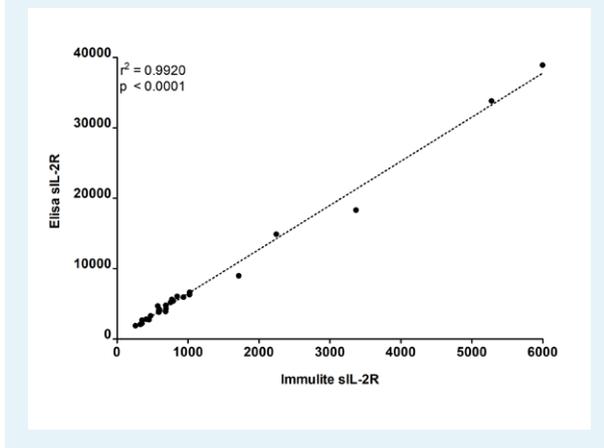
SOLUBLE IL-2 RECEPTOR IN DISEASE AND CLINICAL PRACTICE

Elevated blood sIL-2R levels have been reported in a variety of human diseases, including autoimmune and inflammatory diseases, solid cancers, haematological malignancies, and infections.¹³⁻¹⁶ This clearly indicates the disease aspecificity of elevated sIL-2R. Nevertheless, serum sIL-2R measurement has increased substantially in routine clinical practice over the last decade. Below we discuss laboratory tests for sIL-2R and several conditions where serum sIL-2R measurement can be of clinical use.

Laboratory tests for soluble IL-2 receptor

To date, no gold standard technique and standard reference sera are available for sIL-2R measurement. Current available laboratory tests for sIL-2R comprise enzyme linked immunosorbent assays (ELISA) from different suppliers, but an automated immune assay based on chemoluminescence (CLIA) is also available. A comparison between an ELISA (Diaclone, Besancon Cedex, France) and the automated Immulite chemiluminescent method (Siemens Healthcare, Germany), both commonly used by clinical laboratories in the Netherlands, is shown in figure 3. Although both detection systems report different absolute values in different units (pg/ml vs U/ml) there is perfect correlation between both methods,

Figure 3. Serum levels of sIL-2R as determined by the Immulite chemiluminescent method (Siemens Healthcare, Germany) and ELISA (Diaclone, Besancon Cedex, France). The levels are depicted in different units (Immulinite: U/ml, ELISA: pg/ml) and reveal different absolute values, but with perfect correlation between both methods.



with results differing by a factor of 6-7 in magnitude. Yet, comparison of the Immulite chemiluminescent method with ELISA from another supplier that uses different capture and detection antibodies could result in a different conversion factor. External quality control rounds revealed reproducible results comparing data of the same method.

From a practical point of view, it is important to note that sIL-2R measurements in serum and plasma yield comparable results and that sIL-2R levels remain stable at room temperature over a period of at least three days after sample collection (internal validation, Laboratory Medical Immunology, Department of Immunology, Erasmus MC, University Medical Center Rotterdam, the Netherlands). Also, up to three freeze-thaw cycles do not seem to affect sIL-2R concentration (personal experience).

In healthy individuals, serum sIL-2R levels vary with age, with children and elderly (≥ 65 years) having higher levels than (young) adults.³⁵⁻³⁸ This illustrates that age-related references values are preferable when considering usefulness of serum sIL-2R levels for clinical assessment.

Haemophagocytic lymphohistiocytosis

Haemophagocytic lymphohistiocytosis (HLH) is a complex inflammatory and often very serious disease. Two different HLH forms exist. Primary HLH, also referred to as genetic HLH, has a typical disease onset during infancy or early childhood in individuals with gene mutations that hamper the cytotoxic function of NK cells and T lymphocytes. Secondary HLH tends to occur in older patients in association with another condition, most commonly

malignancy, infection, autoimmune disease, and without an identifiable genetic abnormality.³⁹⁻⁴² Despite the genetic difference between primary HLH and secondary HLH, the clinical manifestations (e.g., fever, hepatosplenomegaly, generalised lymphadenopathy, pancytopenia), and most likely the pathophysiological mechanisms involved, are highly comparable. Although the exact mechanisms are not always clear, it is currently thought that all forms of HLH result from impaired cytotoxic T-lymphocyte and NK-cell functioning. This cellular dysfunction prevents efficient antigen removal with subsequent uncontrolled immune activation with a cytokine storm (including IL-6, IL-18, IFN- γ , TNF- α), uncontrolled macrophage activation, and haemophagocytosis.³⁹⁻⁴³⁻⁴⁵

Serum sIL-2R level is strongly elevated in both primary and secondary HLH and is considered to originate from excessively activated T cells.^{40,45-47} The updated criteria HLH-2004 from the Histiocyte Society included an elevated serum sIL-2R receptor (≥ 2400 U/ml, with a sensitivity of 93% and specificity of 100%) as an additional laboratory diagnostic criterium for paediatric HLH.^{48,49} However, relevant disease controls that most likely would have lowered specificity were lacking in the HLH-2004 study cohort.⁴⁹ A slightly lower sensitivity (89%) for HLH was reported in another paediatric cohort when applying the HLH-2004 serum sIL-2R cut-off value.⁵⁰ Currently the HLH-2004 guidelines contain the standard diagnostic criteria for paediatric HLH. Although developed for paediatric HLH, the HLH-2004 criteria are widely applied to patients with secondary HLH, including adults, as well. There is data to support that the HLH-2004 criteria may be inadequate to accurately diagnose HLH in adults.^{51,52} However, extending HLH-2004 with additional criteria may improve HLH diagnosis in adults, as was for instance, shown for malignancy-associated HLH.⁵² Nevertheless, serum sIL-2R has also been found to display a good to excellent diagnostic performance in diagnosing HLH in adults, with optimal sensitivity (100%) and specificity (72,5%) at a cut-off value of ≥ 2515 U/ml.⁵³ Although this cut-off value is slightly higher than the 2400 U/ml described in the HLH-2004 criteria, the data do demonstrate that a serum sIL-2R level ≤ 2400 U/ml can be helpful in ruling out HLH in adults with high sensitivity (100%). Furthermore, a serum sIL-2R level > 10000 U/ml was found helpful for ruling in HLH in adults with high specificity (93%), but with limited sensitivity (45%).⁵³ Currently, serum sIL-2R is considered a valuable tool in the diagnostic work-up of HLH, yet data on sensitivity and specificity are so far only available from a limited set of studies on paediatric and adult HLH. Establishing a cut-off value for most optimal diagnostic specificity requires further evaluation, especially when one considers the fact that elevated serum sIL-2R levels occur in many diseases, including different types of cancers, infectious conditions,

and autoimmune diseases that can overlap, mimic, or trigger HLH.^{14-16,41,42,54}

There are some reports that suggest that serum sIL-2R might be of use to distinguish the aetiology underlying HLH. For instance, higher levels of serum sIL-2R have been described in malignancy-associated HLH compared to HLH associated with infection or (auto)immune disease.⁵³⁻⁵⁵⁻⁵⁷ However, comparable sIL-2R serum levels between malignancy-associated HLH and EBV-associated HLH have been reported.⁵⁷ Moreover, a higher sIL-2R to ferritin ratio was described in lymphoma-associated HLH as compared to infection-associated HLH and autoimmune disease-associated HLH, but conflicting data exist.⁵³⁻⁵⁵⁻⁵⁶ Likewise, it has been reported that HLH, in the context of (severe)combined immunodeficiency, presents with lower serum sIL-2R levels compared to primary HLH or infection-triggered secondary HLH in infants, which may clearly hamper diagnosing HLH in case of (severe) combined immunodeficiency. Yet, an elevated ratio of serum ferritin/sIL-2R was shown to distinguish HLH in patients with T-cell deficiencies from the other HLH types.⁵⁸ Also, studies reported that primary HLH may present with higher serum sIL-2R levels compared to HLH of other aetiologies, and that the serum sIL-2R/ferritin ratio can distinguish primary HLH from other types of HLH, although data on this is not consistent.^{50,58-60} Thus, although interesting, data to firmly support a role for serum sIL-2R in distinguishing between HLH types and aetiologies are limited and further studies on this are required.

In addition to its application in diagnosing HLH, serum sIL-2R measurement may also provide a tool to monitor disease activity as serum sIL-2R declines with clinical improvement.^{46,53,57,61-63} Alternatively, an increasing serum sIL-2R concentration has been associated with clinical deterioration.⁴⁵⁻⁵³⁻⁵⁴ Moreover, higher initial sIL-2R serum levels (for instance ≥ 10000 U/ml or ≥ 20000 pg/ml) have been reported to be associated with decreased survival compared to HLH patients with lower serum sIL-2R, although data on this is inconclusive.^{53,57,64-66} Altogether these data indicate that serum sIL-2R represents a biomarker useful for at least HLH diagnosis as well as monitoring HLH disease activity and potentially prognosis.

Sarcoidosis

Sarcoidosis is a multisystem granulomatous disease of unknown aetiology presenting with a wide spectrum of clinical manifestations. The natural course of the disease is highly variable and the outcome difficult to predict. Symptom burden is high, and quality of life and social participation are negatively affected. In patients with pulmonary sarcoidosis, treatment is recommended in cases with significant symptoms and/or impaired or deteriorating lung function. The development and

formation of noncaseating granulomas characterises the fundamental abnormality in sarcoidosis, with the lungs, lymph nodes, and skin being the most affected organs.^{67,68} Although granulomas may often resolve spontaneously, pulmonary fibrosis occurs in 10%-15% of patients with sarcoidosis. Granuloma formation is thought to be initiated by CD4+ T lymphocytes that interact with antigen-presenting cells, that become activated and differentiate into Th1 lymphocytes. CD4+ Th1 lymphocytes secrete predominantly IL-2 and IFN- γ and stimulate macrophage TNF- α production, ultimately leading to the characteristic fierce influx of CD4+ T lymphocytes into the involved organs.

A significant correlation was observed between serum soluble IL-2R values and the influx of T lymphocytes into the lungs by Grutters et al., who reported the absolute CD4+ T-lymphocyte numbers in bronchoalveolar lavage in 47 newly diagnosed sarcoidosis patients.⁶⁹ Moreover, higher sIL-2R values were observed in sarcoidosis patients with more advanced and progressive disease, which may predict need for therapy, and high sIL-2R at therapy initiation could serve as a predictor of relapse after infliximab therapy.⁷⁰⁻⁷⁶ Furthermore, sequential measurements of serum sIL-2R could be useful to assess the evolution of disease activity in sarcoidosis. Vorselaars et al. found that decline in serum sIL-2R after six months of treatment with methotrexate correlated with improvement of pulmonary parameters.⁷⁷

In patients with extrapulmonary sarcoidosis, serum sIL-2R was superior to the serum marker, angiotensin-converting enzyme (ACE) when used as screening tool for the detection of intra-ocular sarcoidosis in patients with uveitis.^{78,79} Moreover, Petereit et al. reported that sIL-2R measurements in the cerebrospinal fluid in patients with suspected neurosarcoidosis may help in the diagnostic work-up and may be used to monitor CNS disease activity.⁸⁰ In contrast, in isolated cardiac sarcoidosis compared to non-isolated cardiac sarcoidosis, plasma sIL-2R levels were not elevated.⁸¹ The diagnostic value of sIL-2R for sarcoidosis was confirmed in a recent retrospective cohort study. In total, 189 patients suspected for sarcoidosis were analysed. The sensitivity and specificity of serum sIL-2R for detection of sarcoidosis was 88% and 85%, respectively, superior to ACE (62% and 88%).⁸² In 2015, the diagnostic criteria for sarcoidosis were updated in Japan by the Japanese Society of Sarcoidosis and Other Granulomatous Disorders (JSSOG), with elevated serum sIL-2R replacing negative tuberculin reaction.⁷³ Although considered useful, serum sIL-2R measurement was not included in the recently revised International Workshop on Ocular Sarcoidosis (IWOS) criteria for diagnosing ocular sarcoidosis. The main reason being that it was considered that serum sIL-2R measurement is not (yet) used widely enough in uveitis clinics.⁸³ Nevertheless, the data available

so far do show that serum sIL-2R measurement represents a valuable biomarker in the diagnosis of sarcoidosis as well as for assessment of disease activity and for monitoring treatment efficacy.

Autoimmune disease and other immune-mediated diseases

Increased levels of serum sIL-2R have been described in a variety of autoimmune/immune-mediated diseases as well as other disease conditions associated with immune dysregulation (table 1).^{13-16,78,82,84-102} Of these, rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), juvenile idiopathic arthritis (JIA), adult-onset Still's disease (AOSD), ANCA-associated vasculitis, and IgG4-related disease (IgG4-RD) will be discussed in more detail below.

Rheumatoid arthritis (RA). RA is a chronic inflammatory joint disease that eventually results in destruction of cartilage and bone, severe disability and premature mortality. RA is considered an autoimmune disease associated with the production of autoantibodies, such as anti-cyclic citrullinated peptide (ACPA) and rheumatoid factor (RF).^{103,104} Classification as definite RA, using the ACR/EULAR classification criteria, is based upon the presence of synovitis in at least one joint and a total score of

at least six points (of a possible 10) achieved in four domains. These domains are number and site of involved joints, serological abnormality (ACPA or RF), elevated acute phase response, and at least six weeks of symptom duration.¹⁰⁵

Earlier reports in RA patients revealed conflicting results concerning the correlation between sIL-2R levels and disease activity scores or correlation with other laboratory markers of inflammation.¹⁶ More recently, Kuuliala et al. reported that low sIL-2R levels may be predictive of a rapid response to treatment with infliximab in patients with RA.¹⁰⁶ However, in their cohort, the marker did not identify the patients in remission after 22 weeks. In addition, Steenbergen et al. reported that lower sIL-2R levels were associated with more disease-modifying antirheumatic drug-free sustained remission in RA.²²

Systemic lupus erythematosus (SLE). SLE is a chronic, severely debilitating systemic autoimmune disease characterised by the production of autoantibodies and multi-organ inflammation. SLE is a multifactorial disease that results from complex interactions between susceptibility genes, epigenetic, environmental, hormonal, and immuno-regulatory factors and can present with a wide spectrum of clinical manifestations with unpredictable

Table 1. Examples of immune disorders associated with increased serum soluble interleukin-2 receptor concentrations

Autoimmune/ inflammatory disease	Other immune disease	Associated with immune dysregulation	Other
Adult-onset Still's disease	Allergy	Bipolar disease	Encapsulating peritoneal sclerosis
ANCA-associated vasculitis	Asthma	Complex regional pain syndrome	Graft versus host disease
Atopic dermatitis Behçet's disease Celiac disease Crohn's disease Giant cell arteritis Graves' disease Hemophagocytic lymphohistiocytosis Idiopathic juvenile arthritis IgG4-related disease Multiple sclerosis Myasthenia Gravis Myositis Non-ANCA vasculitis Non-infectious uveitis Rheumatoid arthritis Sarcoidosis Sjögren's syndrome Systemic lupus erythematosus Systemic sclerosis Type-1 diabetes	Granulomatous CVID	Obesity	Transplant rejection

ANCA = antineutrophil cytoplasmic antibodies; CVID = common variable immunodeficiency disorder

relapse-remitting course.^{107,108} SLE may involve almost all organs and tissues. Clinical manifestations may include fatigue, mucocutaneous lesions, renal involvement, arthritis, haematological abnormalities, serositis and fever. Newly developed 2019 EULAR/ACR classification criteria for SLE include positive antinuclear antibody (ANA) as entry criterion, followed by weighed criteria grouped in seven clinical domains (constitutional, haematological, neuropsychiatric, mucocutaneous, serosal, musculoskeletal, renal) and three immunological domains (antiphospholipid antibodies, complement proteins, and SLE-specific antibodies). Patients fulfil classification for SLE when accumulated ≥ 10 points.¹⁰⁹

Elevated blood levels of sIL-2R occur in SLE and have been reported to precede major disease exacerbations.¹¹⁰⁻¹¹² Also, significantly higher sIL-2R values are found in SLE patients with lupus nephritis compared to SLE patients without nephritis, and sIL-2R levels decline after treatment.^{113,114} Recently, Zhang et al. reported that SLE patients in the group with high sIL-2R values had significantly more lupus nephritis, arthritis, and vasculitis compared to SLE patients in the group with low sIL-2R values. Moreover, high sIL-2R values were significantly associated with laboratory parameters of renal impairment and Systemic Lupus Erythematosus Disease Activity Index 2000 (SLEDAI-2K).¹¹⁵ Together, these data indicate that sIL-2R may be a useful biomarker for the assessment of SLE disease activity and might be used as early indicator of renal involvement. As T lymphocytes play a central role in most rheumatic diseases, it would be of interest to establish correlations between sIL-2R values and disease activity scores in other connective tissue diseases, e.g., systemic sclerosis and Sjögren's disease.

Juvenile idiopathic arthritis (JIA). JIA is a heterogeneous group of diseases characterised by arthritis of unknown origin with onset before age of 16 years. The current International League of Associations for Rheumatology (ILAR) classification criteria for JIA were developed by consensus and not formally validated.^{116,117} The classification criteria recognise six mutually exclusive categories defined in clinical and laboratory measures: systemic arthritis, oligoarthritis (persistent or extended), polyarthritis rheumatoid factor (RF)-positive, polyarthritis RF-negative, enthesitis-related arthritis, psoriatic arthritis, and a seventh category, undifferentiated arthritis. Recently, an initiative was started to provide new evidence-based classification of JIA using a formal process to validate preliminary criteria.¹¹⁸

Elevated sIL-2R levels were found in patients with clinically active JIA compared to controls,¹¹⁹⁻¹²¹ and sIL-2R levels correlated significantly with pannus thickness and joint count.^{120,121} Furthermore, in addition to other disease characteristics, treatment-refractory disease

course may be associated with a higher sIL-2R level.¹²² Finally, macrophage activation syndrome (MAS) has been increasingly recognised in association with rheumatic diseases, most commonly in systemic JIA. The clinical features of MAS in JIA are similar to HLH and include high, non-remitting fever, generalised lymphadenopathy, hepatosplenomegaly, central nervous system dysfunction, and haemorrhagic manifestations and can result in multi-organ failure.¹²³ Recent reports suggest that (subclinical) MAS in systemic JIA may even occur in 30-40% of patients.^{124,125} The utility of serum sIL-2R for JIA will probably vary per clinical phenotype, but, serum sIL-2R may be a promising marker for disease activity in systemic JIA, especially when associated with MAS.¹²⁴

Adult-onset Still's disease (AOSD). AOSD is a rare systemic inflammatory disorder characterised by high fever that typically spikes once or twice daily, a transient salmon pink skin rash (mostly on the proximal limbs and trunk) that occurs along with the fever spikes, arthritis and arthralgia involving predominantly the wrists, knees and ankles, and frequently a sore throat; as well as less frequent symptoms including myalgias, lymphadenopathies, splenomegaly, hepatomegaly, pleurisy, pericarditis, weight loss, and abdominal pain.^{126,127} The pathophysiology of AOSD is mainly unknown, but involvement of a pro-inflammatory cascade that can be triggered by infectious agents, solid cancers, or lymphomas in genetically predisposed individuals has been proposed.^{126,127} AOSD shares several phenotypic characteristics with systemic JIA, including daily recurring fever, salmon-coloured skin rash and polyarthritis, and is considered to represent the adult counterpart of systemic JIA or even a continuum of a single disease entity.^{127,128}

No definitive diagnostic tool for AOSD exists and its diagnosis is based on extensively excluding diseases with comparable clinical presentation (including, for example, viral and bacterial infections, SLE, RA, myositis, systemic vasculitis, autoinflammatory diseases, sarcoidosis, malignancies), for which several diagnostic criteria sets have been developed.¹²⁷ Laboratory findings reflect the (non-specific) systemic inflammatory nature of the disease and increased erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) levels, neutrophilic leucocytosis ($> 80\%$ polymorphonuclear leucocytes), anaemia and thrombocytosis are common findings. Also, serum ferritin is highly elevated in AOSD, while glycosylated serum ferritin is typically low.^{126,127} Neutrophil and macrophage activation are at the centre of the immunopathogenesis of AOSD and macrophage-derived pro-inflammatory cytokines such as the interleukin-1 family members IL-1 β and IL-18, as well as IL-6, are centrally involved.^{126,127} AOSD is also strongly associated with secondary HLH and cytotoxic functions

of NK cells are diminished in active AOSD.^{126,127,129,130} Also, increased IFN- γ -producing Th1 lymphocytes have been detected in peripheral blood and pathological tissues from patients with active untreated AOSD, which likely contributes to further macrophage activation and cell-mediated immunity.^{126,127,131} In addition, circulating Th17 lymphocytes were found elevated in patients with active untreated AOSD.¹³² Frequencies of circulating Th1 and Th17 lymphocytes correlated significantly with clinical activity, and serum IL-18 and ferritin levels in active untreated AOSD, and all these laboratory parameters declined with clinical remission upon treatment.^{131,132}

Only limited data on serum sIL-2R in AOSD is available, yet increased levels are detected in active AOSD, thus further supporting T-lymphocyte involvement in this disease.^{131,133-137} Although AOSD may not display differences in serum sIL-2R levels compared to other diseases, serum sIL-2R level may represent a potential biomarker for monitoring AOSD disease activity and treatment response.¹³⁶ Serum sIL-2R levels have been reported to correlate with AOSD disease activity.^{133,135,137} Moreover, Fuji et al. reported that serum sIL-2R levels were higher in the subgroup of AOSD patients with chronic articular disease, suggesting that serum sIL-2R levels may distinguish between different AOSD disease patterns.¹³⁵ Also, several studies demonstrated that upon treatment, disease remission was associated with a strong decline in serum sIL-2R levels, along with decreases in other laboratory parameters such as ESR, CRP, ferritin, and IL-18 levels.^{131,134,135,137,138} In contrast, a rise in serum sIL-2R level can occur in case of disease recurrence.¹³⁷

Altogether, the data available so far suggest that serum sIL-2R can be considered as an additional biomarker to monitor AOSD disease activity and therapeutic response.

ANCA-associated vasculitis (AAV). AAV is a necrotising vasculitis that predominantly affects small vessels and is associated with antineutrophil cytoplasmic antibodies (ANCA) specific for myeloperoxidase (MPO-ANCA) or proteinase 3 (PR3-ANCA). The major clinicopathological variants of AAV are microscopic polyangiitis (MPA), granulomatosis with polyangiitis (GPA), and eosinophilic granulomatosis with polyangiitis (EGPA). Besides necrotising vasculitis, GPA and EGPA show necrotising granulomatous inflammation often involving the respiratory tract. Moreover, EGPA is associated with asthma and eosinophilia.¹³⁹ Clinical manifestations that suggest the diagnosis of AAV, when there is no other obvious cause, and indicate ANCA testing include: (rapid progressive) glomerulonephritis, pulmonary haemorrhage, cutaneous vasculitis with systemic features, multiple lung nodules, chronic destructive disease of the upper airways, long-standing sinusitis or otitis, subglottic tracheal stenosis, peripheral neuropathy, retro-orbital mass, and scleritis.¹⁴⁰

Elevated levels of sIL-2R have been detected in the sera of patients with GPA and MPA. Moreover, sIL-2R levels correlated with disease activity at diagnosis and differed between limited and generalised and between active and inactive disease.¹⁴¹⁻¹⁴⁴ In addition, positive ANCA serology during follow up was associated with sIL-2R levels. At 18 and at 24 months after diagnosis, higher levels of sIL-2R were found in ANCA-positive patients compared to patients negative for ANCA.¹⁴² Analogous to sIL-2R as marker for T-cell activation, soluble CD163 (sCD163) is considered a systemic marker of macrophage activation. CD163 is abundantly expressed by tissue macrophages and is shed from the macrophage surface under inflammatory conditions by ADAM17, the enzyme that also releases TNF into the circulation.^{145,146} It has been reported that urinary sCD163 levels may reflect active glomerular inflammation.¹⁴⁷ Dekkema et al. found that measurement of serum sIL-2R, in addition to urinary sIL-2R, complements urinary sCD163 in the detection of active renal vasculitis in AAV patients and that serum and urinary sIL-2R are significantly higher during active renal disease and decline upon remission.¹⁴⁸

Taken together, these data show that sIL-2R may be a valuable biomarker for assessment of disease activity and for monitoring treatment, and seems to reflect the central role of the T-lymphocyte-driven immune response in ANCA-associated vasculitis.

IgG4-related disease (IgG4-RD). IgG4-RD is a fibroinflammatory disease that can involve various organs, including the lungs, thyroid, lymph nodes, orbital tissue, kidneys, salivary and lacrimal glands, aorta, pancreas, and skin. IgG4-RD is characterised by accumulation of IgG4-producing plasma cells at affected sites, along with the formation of storiform fibrotic lesions.¹⁴⁹ Serum IgG4 levels are elevated in the majority of patients with IgG4-RD, yet IgG4 elevation is not fully sensitive or specific for diagnosing IgG4-RD.¹⁴⁹ Moreover, serum IgG4 levels may not always accurately reflect disease activity.^{150,151} Therefore, additional blood biomarkers for improved diagnosis and evaluation of IgG4-RD disease activity are still needed.^{152,153}

To date, only a limited number of studies have explored serum sIL-2R in relation to IgG4-RD. These studies report elevated serum sIL-2R levels in IgG4-RD, a positive correlation between serum sIL-2R levels with the number of affected organs as well as disease activity.^{90,154,155} Moreover, these studies reported a decline of serum sIL-2R levels after treatment.^{90,154,155} In addition, serum sIL-2R levels have been reported to display high accuracy in predicting an individual glucocorticoid requirement.¹⁵⁵ These data indicate that serum sIL-2R level may be a valuable biomarker for evaluating disease activity and treatment response in IgG4-RD. However, additional

(prospective) studies are needed, especially with regard to treatment stratification, monitoring treatment response, as well as sensitivity and specificity of sIL-2R in the context of IgG4-RD diagnosis.

Cancer and treatment

Increased serum sIL-2R levels have been found in a variety of malignancies, mostly haematopoietic malignancies but also solid cancers.¹³⁻¹⁶ These elevated sIL-2R levels most likely derive from the malignant cells, although the host cellular immune response likely contributes to the generation of sIL-2R as well.¹³⁻¹⁵

Immune checkpoint inhibitor treatment is a rapidly expanding field within oncology. High serum sIL-2R in metastatic melanoma patients before initiation of ipilimumab (anti-CTLA-4) treatment has been associated with treatment resistance, most likely by IL-2 sequestration.¹⁵⁶ Despite the anti-tumour efficacy of immune checkpoint inhibitor treatment, secondary development or worsening of autoimmune/inflammatory disorders is commonly observed in cancer patients upon such treatment.¹⁵⁷ Measurement of serum sIL-2R prior and during therapy, potentially along with other cytokines such as IFN- γ , IL-17, and IL-10, may, in the future, prove

a valuable tool for monitoring excessive/uncontrolled immune activation and to predict the development of immune-related adverse events in cancer patients treated with immune checkpoint inhibitors.¹⁵⁷

CONCLUDING REMARKS

The clinical utility of sIL-2R, as reviewed above, lies particularly in evaluating disease activity in a variety of immune-mediated diseases and may add in the diagnostic work-up of especially HLH and sarcoidosis. As sIL-2R level reflects activation status of the T-lymphocyte compartment, its disease specific value is limited. However, in immune-mediated diseases where T-lymphocyte responses play a central role in the pathophysiology, sIL-2R might be a valuable biomarker for predicting or monitoring the efficacy of immune suppressive therapies.

DISCLOSURE

All authors declare no conflicts of interest. No funding or financial support was received.

REFERENCES

- Boyman O, Sprent J. The role of interleukin-2 during homeostasis and activation of the immune system. *Nat Rev Immunol.* 2012;12:180-90.
- Setoguchi R, Hori S, Takahashi T, Sakaguchi S. Homeostatic maintenance of natural Foxp3(+) CD25(+) CD4(+) regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. *J Exp Med.* 2005;201:723-35.
- Spolski R, Li P, Leonard WJ. Biology and regulation of IL-2: from molecular mechanisms to human therapy. *Nat Rev Immunol.* 2018;18:648-59.
- Boyman O, Kovar M, Rubinstein MP, Surh CD, Sprent J. Selective stimulation of T cell subsets with antibody-cytokine immune complexes. *Science.* 2006;311:1924-7.
- Kim HP, Imbert J, Leonard WJ. Both integrated and differential regulation of components of the IL-2/IL-2 receptor system. *Cytokine Growth Factor Rev.* 2006;17:349-66.
- Rubin LA, Kurman CC, Fritz ME, et al. Soluble interleukin 2 receptors are released from activated human lymphoid cells in vitro. *J Immunol.* 1985;135:3172-7.
- Lai KN, Leung JC, Lai FM. Soluble interleukin 2 receptor release, interleukin 2 production, and interleukin 2 receptor expression in activated T-lymphocytes in vitro. *Pathology.* 1991;23:224-8.
- Pedersen AE, Lauritsen JP. CD25 shedding by human natural occurring CD4+CD25+ regulatory T cells does not inhibit the action of IL-2. *Scand J Immunol.* 2009;70:40-3.
- Bank U, Reinhold D, Schneemilch C, Kunz D, Synowitz HJ, Ansoerge S. Selective proteolytic cleavage of IL-2 receptor and IL-6 receptor ligand binding chains by neutrophil-derived serine proteases at foci of inflammation. *J Interferon Cytokine Res.* 1999;19:1277-87.
- Schulz O, Sewell HF, Shakib F. Proteolytic cleavage of CD25, the alpha subunit of the human T cell interleukin 2 receptor, by Der p 1, a major mite allergen with cysteine protease activity. *J Exp Med.* 1998;187:271-5.
- Sheu BC, Hsu SM, Ho HN, Lien HC, Huang SC, Lin RH. A novel role of metalloproteinase in cancer-mediated immunosuppression. *Cancer Res.* 2001;61:237-42.
- Rubin LA, Galli F, Greene WC, Nelson DL, Jay G. The molecular basis for the generation of the human soluble interleukin 2 receptor. *Cytokine.* 1990;2:330-6.
- Bien E, Balcerska A. Serum soluble interleukin 2 receptor alpha in human cancer of adults and children: a review. *Biomarkers.* 2008;13:1-26.
- Caruso C, Candore G, Cigna D, Colucci AT, Modica MA. Biological significance of soluble IL-2 receptor. *Mediators Inflamm.* 1993;2:3-21.
- Rubin LA, Nelson DL. The soluble interleukin-2 receptor: biology, function, and clinical application. *Ann Intern Med.* 1990;113:619-27.
- Witkowska AM. On the role of sIL-2R measurements in rheumatoid arthritis and cancers. *Mediators Inflamm.* 2005;2005:121-30.
- Brand OJ, Lowe CE, Heward JM, et al. Association of the interleukin-2 receptor alpha (IL-2Ralpha)/CD25 gene region with Graves' disease using a multilocus test and tag SNPs. *Clin Endocrinol (Oxf).* 2007;66:508-12.
- Carr EJ, Clatworthy MR, Lowe CE, et al. Contrasting genetic association of IL2RA with SLE and ANCA-associated vasculitis. *BMC Med Genet.* 2009;10:22.
- International Multiple Sclerosis Genetics C, Hafler DA, Compston A, et al. Risk alleles for multiple sclerosis identified by a genome-wide study. *N Engl J Med.* 2007;357:851-62.
- Lowe CE, Cooper JD, Brusko T, et al. Large-scale genetic fine mapping and genotype-phenotype associations implicate polymorphism in the IL2RA region in type 1 diabetes. *Nat Genet.* 2007;39:1074-82.
- Maier LM, Anderson DE, Severson CA, et al. Soluble IL-2RA levels in multiple sclerosis subjects and the effect of soluble IL-2RA on immune responses. *J Immunol.* 2009;182:1541-7.
- van Steenberg HW, van Nies JA, Ruyssen-Witrand A, et al. IL2RA is associated with persistence of rheumatoid arthritis. *Arthritis Res Ther.* 2015;17:244.
- Nelson DL, Rubin LA, Kurman CC, Fritz ME, Boutin B. An analysis of the cellular requirements for the production of soluble interleukin-2 receptors in vitro. *J Clin Immunol.* 1986;6:114-20.

24. Holter W, Goldman CK, Casabo L, Nelson DL, Greene WC, Waldmann TA. Expression of functional IL 2 receptors by lipopolysaccharide and interferon-gamma stimulated human monocytes. *J Immunol.* 1987;138:2917-22.
25. Kniep EM, Strelow I, Lohmann-Matthes ML. The monocyte interleukin-2 receptor light chain: production of cell-associated and soluble interleukin-2 receptor by monocytes. *Immunology.* 1992;75:299-304.
26. Valitutti S, Carbone A, Castellino F, et al. The expression of functional IL-2 receptor on activated macrophages depends on the stimulus applied. *Immunology.* 1989;67:44-50.
27. von Bergwelt-Baildon MS, Popov A, Saric T, et al. CD25 and indoleamine 2,3-dioxygenase are up-regulated by prostaglandin E2 and expressed by tumor-associated dendritic cells in vivo: additional mechanisms of T-cell inhibition. *Blood.* 2006;108:228-37.
28. Rubin LA, Jay G, Nelson DL. The released interleukin 2 receptor binds interleukin 2 efficiently. *J Immunol.* 1986;137:3841-4.
29. Lindqvist CA, Christiansson LH, Simonsson B, Enblad G, Olsson-Stromberg U, Loskog AS. T regulatory cells control T-cell proliferation partly by the release of soluble CD25 in patients with B-cell malignancies. *Immunology.* 2010;131:371-6.
30. Rubinstein MP, Kovar M, Purton JF, et al. Converting IL-15 to a superagonist by binding to soluble IL-15R[alpha]. *Proc Natl Acad Sci U S A.* 2006;103:9166-71.
31. Kobayashi H, Tagaya Y, Han ES, et al. Use of an antibody against the soluble interleukin 2 receptor alpha subunit can modulate the stability and biodistribution of interleukin-2. *Cytokine.* 1999;11:1065-75.
32. Vanmaris RMM, Rijkers GT. Biological role of the soluble interleukin-2 receptor in sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis.* 2017;34:122-9.
33. Yang ZZ, Grote DM, Ziesmer SC, et al. Soluble IL-2Ralpha facilitates IL-2-mediated immune responses and predicts reduced survival in follicular B-cell non-Hodgkin lymphoma. *Blood.* 2011;118:2809-20.
34. Russell SE, Moore AC, Fallon PG, Walsh PT. Soluble IL-2Ralpha (sCD25) exacerbates autoimmunity and enhances the development of Th17 responses in mice. *PLoS One.* 2012;7:e47748.
35. Filipovich AH. Hemophagocytic lymphohistiocytosis and other hemophagocytic disorders. *Immunol Allergy Clin North Am.* 2008;28:293-313, viii.
36. Gotoh Y, Okamoto Y, Uemura O, et al. Determination of age-related changes in human soluble interleukin 2 receptor in body fluids of normal subjects as a control value against disease states. *Clin Chim Acta.* 1999;289:89-97.
37. Manoussakis MN, Stavropoulos ED, Germanidis GS, et al. Soluble interleukin-2 receptors and autoantibodies in the serum of healthy elderly individuals. *Autoimmunity.* 1990;7:129-37.
38. Sack U, Burkhardt U, Borte M, Schadlich H, Berg K, Emmrich F. Age-dependent levels of select immunological mediators in sera of healthy children. *Clin Diagn Lab Immunol* 1998;5:28-32.
39. Rosado FG, Kim AS. Hemophagocytic lymphohistiocytosis: an update on diagnosis and pathogenesis. *Am J Clin Pathol.* 2013;139:713-27.
40. Ramos-Casals M, Brito-Zeron P, Lopez-Guillermo A, Khamashta MA, Bosch X. Adult haemophagocytic syndrome. *Lancet.* 2014;383:1503-16.
41. Parikh SA, Kapoor P, Letendre L, Kumar S, Wolanskyj AP. Prognostic factors and outcomes of adults with hemophagocytic lymphohistiocytosis. *Mayo Clin Proc.* 2014;89:484-92.
42. Riviere S, Galicier L, Coppo P, et al. Reactive hemophagocytic syndrome in adults: a retrospective analysis of 162 patients. *Am J Med.* 2014;127:1118-25.
43. Henter JL, Elinder G, Soder O, Hansson M, Andersson B, Andersson U. Hypercytokinemia in familial hemophagocytic lymphohistiocytosis. *Blood.* 1991;78:2918-22.
44. Schaer DJ, Schleiffenbaum B, Kurrer M, et al. Soluble hemoglobin-haptoglobin scavenger receptor CD163 as a lineage-specific marker in the reactive hemophagocytic syndrome. *Eur J Haematol.* 2005;74:6-10.
45. Zondag TC, Roxk C, van Lom K, et al. Cytokine and viral load kinetics in human herpesvirus 8-associated multicentric Castlemans disease complicated by hemophagocytic lymphohistiocytosis. *Int J Hematol.* 2016;103:469-72.
46. Komp DM, McNamara J, Buckley P. Elevated soluble interleukin-2 receptor in childhood hemophagocytic histiocytic syndromes. *Blood.* 1989;73:2128-32.
47. Lin M, Park S, Hayden A, et al. Clinical utility of soluble interleukin-2 receptor in hemophagocytic syndromes: a systematic scoping review. *Ann Hematol.* 2017;96:1241-51.
48. Henter JL, Horne A, Arico M, et al. HLH-2004: Diagnostic and therapeutic guidelines for hemophagocytic lymphohistiocytosis. *Pediatr Blood Cancer.* 2007;48:124-31.
49. Lehmborg K, Ehl S. Diagnostic evaluation of patients with suspected haemophagocytic lymphohistiocytosis. *Br J Haematol.* 2013;160:275-87.
50. Lehmborg K, Pink I, Eulenburg C, Beutel K, Maul-Pavicic A, Janka G. Differentiating macrophage activation syndrome in systemic juvenile idiopathic arthritis from other forms of hemophagocytic lymphohistiocytosis. *J Pediatr.* 2013;162:1245-51.
51. Raschke RA, Garcia-Orr R. Hemophagocytic lymphohistiocytosis: a potentially underrecognized association with systemic inflammatory response syndrome, severe sepsis, and septic shock in adults. *Chest.* 2011;140:933-8.
52. Tamamyian GN, Kantarjian HM, Ning J, et al. Malignancy-associated hemophagocytic lymphohistiocytosis in adults: Relation to hemophagocytosis, characteristics, and outcomes. *Cancer.* 2016;122:2857-66.
53. Hayden A, Lin M, Park S, et al. Soluble interleukin-2 receptor is a sensitive diagnostic test in adult HLH. *Blood Adv.* 2017;1:2529-34.
54. Schram AM, Berliner N. How I treat hemophagocytic lymphohistiocytosis in the adult patient. *Blood.* 2015;125:2908-14.
55. Tabata C, Tabata R. Possible prediction of underlying lymphoma by high sIL-2R/ferritin ratio in hemophagocytic syndrome. *Ann Hematol.* 2012;91:63-71.
56. Tsuji T, Hirano T, Yamasaki H, Tsuji M, Tsuda H. A high sIL-2R/ferritin ratio is a useful marker for the diagnosis of lymphoma-associated hemophagocytic syndrome. *Ann Hematol.* 2014;93:821-6.
57. Zhang L, Zhang S, Xu J, et al. Significance of soluble interleukin-2 receptor in patients with hemophagocytic lymphohistiocytosis. *Leuk Lymphoma.* 2011;52:1360-2.
58. Bode SF, Ammann S, Al-Herz W, et al. The syndrome of hemophagocytic lymphohistiocytosis in primary immunodeficiencies: implications for differential diagnosis and pathogenesis. *Haematologica.* 2015;100:978-88.
59. Yasumi T, Hori M, Hiejima E, et al. Laboratory parameters identify familial haemophagocytic lymphohistiocytosis from other forms of paediatric haemophagocytosis. *Br J Haematol.* 2015;170:532-8.
60. Chen Y, Wang Z, Luo Z, Zhao N, Yang S, Tang Y. Comparison of Th1/Th2 cytokine profiles between primary and secondary haemophagocytic lymphohistiocytosis. *Ital J Pediatr.* 2016;42:50.
61. Mellor-Heineke S, Villanueva J, Jordan MB, et al. Elevated Granzyme B in Cytotoxic Lymphocytes is a Signature of Immune Activation in Hemophagocytic Lymphohistiocytosis. *Front Immunol.* 2013;4:72.
62. Faguer S, Vergez F, Peres M, et al. Tocilizumab added to conventional therapy reverses both the cytokine profile and CD8+Granzyme+ T-cells/NK cells expansion in refractory hemophagocytic lymphohistiocytosis. *Hematol Oncol.* 2016;34:55-7.
63. Ahmed A, Merrill SA, Alsawah F, et al. Ruxolitinib in adult patients with secondary haemophagocytic lymphohistiocytosis: an open-label, single-centre, pilot trial. *Lancet Haematol.* 2019;6:e630-e7.
64. Fujiwara F, Hibi S, Imashuku S. Hypercytokinemia in hemophagocytic syndrome. *Am J Pediatr Hematol Oncol.* 1993;15:92-8.
65. Imashuku S, Hibi S, Sako M, et al. Soluble interleukin-2 receptor: a useful prognostic factor for patients with hemophagocytic lymphohistiocytosis. *Blood.* 1995;86:4706-7.
66. Imashuku S, Ikushima S, Esumi N, Todo S, Saito M. Serum Levels of Interferon-gamma, Cytotoxic Factor and Soluble Interleukin-2 Receptor in Childhood Hemophagocytic Syndromes. *Leuk Lymphoma.* 1991;3:287-92.
67. Grunewald J, Grutters JC, Arkema EV, Saketkoo LA, Moller DR, Muller-Quernheim J. Sarcoidosis. *Nat Rev Dis Primers.* 2019;5:45.
68. Iannuzzi MC, Rybicki BA, Teirstein AS. Sarcoidosis. *N Engl J Med.* 2007;357:2153-65.
69. Grutters JC, Fellrath JM, Mulder L, Janssen R, van den Bosch JM, van Velzen-Blad H. Serum soluble interleukin-2 receptor measurement in patients with sarcoidosis: a clinical evaluation. *Chest.* 2003;124:186-95.
70. Miyoshi S, Hamada H, Kadowaki T, et al. Comparative evaluation of serum markers in pulmonary sarcoidosis. *Chest* 2010;137:1391-7.

71. Rothkrantz-Kos S, van Dieijen-Visser MP, Mulder PG, Drent M. Potential usefulness of inflammatory markers to monitor respiratory functional impairment in sarcoidosis. *Clin Chem.* 2003;49:1510-7.
72. Schimmelpennink MC, Vorselaars ADM, van Beek FT, et al. Efficacy and safety of infliximab biosimilar Inflectra((R)) in severe sarcoidosis. *Respir Med.* 2018;138S:57-S13.
73. Thi Hong Nguyen C, Kambe N, Kishimoto I, Ueda-Hayakawa I, Okamoto H. Serum soluble interleukin-2 receptor level is more sensitive than angiotensin-converting enzyme or lysozyme for diagnosis of sarcoidosis and may be a marker of multiple organ involvement. *J Dermatol* 2017;44:789-97.
74. Vorselaars AD, Verwoerd A, van Moorsel CH, Keijsers RG, Rijkers GT, Grutters JC. Prediction of relapse after discontinuation of infliximab therapy in severe sarcoidosis. *Eur Respir J.* 2014;43:602-9.
75. Ziegenhagen MW, Benner UK, Zissel G, Zabel P, Schlaak M, Muller-Quernheim J. Sarcoidosis: TNF-alpha release from alveolar macrophages and serum level of sIL-2R are prognostic markers. *Am J Respir Crit Care Med.* 1997;156:1586-92.
76. Ziegenhagen MW, Rothe ME, Schlaak M, Muller-Quernheim J. Bronchoalveolar and serological parameters reflecting the severity of sarcoidosis. *Eur Respir J.* 2003;21:407-13.
77. Vorselaars AD, van Moorsel CH, Zanen P, et al. ACE and sIL-2R correlate with lung function improvement in sarcoidosis during methotrexate therapy. *Respir Med.* 2015;109:279-85.
78. Groen-Hakan F, Eurelings L, ten Berge JC, et al. Diagnostic Value of Serum-Soluble Interleukin 2 Receptor Levels vs Angiotensin-Converting Enzyme in Patients With Sarcoidosis-Associated Uveitis. *JAMA Ophthalmol.* 2017;135:1352-8.
79. Gundlach E, Hoffmann MM, Prasse A, Heinzlmann S, Ness T. Interleukin-2 Receptor and Angiotensin-Converting Enzyme as Markers for Ocular Sarcoidosis. *PLoS One.* 2016;11:e0147258.
80. Petereit HF, Reske D, Tumani H, et al. Soluble CSF interleukin 2 receptor as indicator of neurosarcoidosis. *J Neurol.* 2010;257:1855-63.
81. Kiko T, Yoshihisa A, Kanno Y, et al. A Multiple Biomarker Approach in Patients with Cardiac Sarcoidosis. *Int Heart J.* 2018;59:996-1001.
82. Eurelings LEM, Miedema JR, Dalm V, et al. Sensitivity and specificity of serum soluble interleukin-2 receptor for diagnosing sarcoidosis in a population of patients suspected of sarcoidosis. *PLoS One.* 2019;14:e0223897.
83. Mochizuki M, Smith JR, Takase H, et al. Revised criteria of International Workshop on Ocular Sarcoidosis (IWOS) for the diagnosis of ocular sarcoidosis. *Br J Ophthalmol.* 2019;103:1418-22.
84. Alpsoy E, Cayirli C, Er H, Yilmaz E. The levels of plasma interleukin-2 and soluble interleukin-2R in Behcet's disease: a marker of disease activity. *J Dermatol.* 1998;25:513-6.
85. Betjes MG, Habib MS, Struijk DG, et al. Encapsulating peritoneal sclerosis is associated with T-cell activation. *Nephrol Dial Transplant.* 2015;30:1568-76.
86. Bharwani KD, Dik WA, Dirckx M, Huygen F. Highlighting the Role of Biomarkers of Inflammation in the Diagnosis and Management of Complex Regional Pain Syndrome. *Mol Diagn Ther.* 2019;23:615-26.
87. Bharwani KD, Dirckx M, Stronks DL, Dik WA, Schreurs MWJ, Huygen F. Elevated Plasma Levels of sIL-2R in Complex Regional Pain Syndrome: A Pathogenic Role for T-Lymphocytes? *Mediators Inflamm.* 2017;2017:2764261.
88. Breunis MN, Kupka RW, Nolen WA, et al. High numbers of circulating activated T cells and raised levels of serum IL-2 receptor in bipolar disorder. *Biol Psychiatry.* 2003;53:157-65.
89. Del Vecchio GC, Penza R, Altomare M, et al. Cytokine pattern and endothelium damage markers in Henoch-Schonlein purpura. *Immunopharmacol Immunotoxicol.* 2008;30:623-9.
90. Karim AF, Eurelings LEM, Bansie RD, van Hagen PM, van Laar JAM, Dik WA. Soluble Interleukin-2 Receptor: A Potential Marker for Monitoring Disease Activity in IgG4-Related Disease. *Mediators Inflamm.* 2018;2018:6103064.
91. Kowal K, Pampuch A, Kowal-Bielecka O, Iacoviello L, Bodzenta-Lukaszyk A. Soluble CD40 ligand in asthma patients during allergen challenge. *J Thromb Haemost.* 2006;4:2718-20.
92. Nakamura H, Komatsu K, Ayaki M, et al. Serum levels of soluble IL-2 receptor, IL-12, IL-18, and IFN-gamma in patients with acute graft-versus-host disease after allogeneic bone marrow transplantation. *J Allergy Clin Immunol.* 2000;106:S45-50.
93. Netea MG, Hancu N. Increased soluble interleukin-2 receptor concentrations in patients with insulin-dependent diabetes mellitus. *Diabet Med.* 1997;14:168.
94. Nishioka A, Tsunoda S, Abe T, et al. Serum neopterin as well as ferritin, soluble interleukin-2 receptor, KL-6 and anti-MDA5 antibody titer provide markers of the response to therapy in patients with interstitial lung disease complicating anti-MDA5 antibody-positive dermatomyositis. *Mod Rheumatol.* 2019;29:814-20.
95. Ohashi Y, Tanaka A, Kakinoki Y, et al. Serum level of soluble interleukin-2 receptor in patients with seasonal allergic rhinitis. *Scand J Immunol.* 1997;45:315-21.
96. Stelmach I, Jerzynska J, Kuna P. A randomized, double-blind trial of the effect of treatment with montelukast on bronchial hyperresponsiveness and serum eosinophilic cationic protein (ECP), soluble interleukin 2 receptor (sIL-2R), IL-4, and soluble intercellular adhesion molecule 1 (sICAM-1) in children with asthma. *J Allergy Clin Immunol.* 2002;109:257-63.
97. Thijs JL, Drylewicz J, Fiechter R, et al. EASI p-EASI: Utilizing a combination of serum biomarkers offers an objective measurement tool for disease severity in atopic dermatitis patients. *J Allergy Clin Immunol.* 2017;140:1703-5.
98. Tournadre A, Dubost JJ, Soubrier M, et al. Soluble IL-2 receptor: a biomarker for assessing myositis activity. *Dis Markers.* 2014;2014:472624.
99. van der Zalm LJB, van der Valk ES, Wester VL, et al. Obesity-associated T-cell and macrophage activation is partly reversible by lifestyle intervention. *Int J Obes* 2020;in press.
100. Vitale J, Convers KD, Goretzke S, et al. Serum IL-12 and soluble IL-2 receptor levels as possible biomarkers of granulomatous and lymphocytic interstitial lung disease in common variable immunodeficiency: a case report. *J Allergy Clin Immunol Pract.* 2015;3:273-6.
101. Wolf RE, Baethge BA. Interleukin-1 alpha, interleukin-2, and soluble interleukin-2 receptors in polymyositis. *Arthritis Rheum.* 1990;33:1007-14.
102. Yoshizawa Y, Nomaguchi H, Izaki S, Kitamura K. Serum cytokine levels in atopic dermatitis. *Clin Exp Dermatol.* 2002;27:225-9.
103. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med.* 2011;365:2205-19.
104. Nielen MM, van Schaardenburg D, Reesink HW, et al. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis Rheum.* 2004;50:380-6.
105. Aletaha D, Neogi T, Silman AJ, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Ann Rheum Dis.* 2010;69:1580-8.
106. Kuuliala A, Nissinen R, Kautiainen H, Repo H, Leirisalo-Repo M. Low circulating soluble interleukin 2 receptor level predicts rapid response in patients with refractory rheumatoid arthritis treated with infliximab. *Ann Rheum Dis.* 2006;65:26-9.
107. Tsokos GC. Systemic lupus erythematosus. *N Engl J Med.* 2011;365:2110-21.
108. Choi J, Kim ST, Craft J. The pathogenesis of systemic lupus erythematosus-an update. *Curr Opin Immunol.* 2012;24:651-7.
109. Aringer M, Costenbader K, Daikh D, et al. 2019 European League Against Rheumatism/American College of Rheumatology classification criteria for systemic lupus erythematosus. *Ann Rheum Dis.* 2019;78:1151-9.
110. ter Borg EJ, Horst G, Limburg PC, Kallenberg CG. Changes in plasma levels of interleukin-2 receptor in relation to disease exacerbations and levels of anti-dsDNA and complement in systemic lupus erythematosus. *Clin Exp Immunol.* 1990;82:21-6.
111. Spronk PE, ter Borg EJ, Huitema MG, Limburg PC, Kallenberg CG. Changes in levels of soluble T-cell activation markers, sIL-2R, sCD4 and sCD8, in relation to disease exacerbations in patients with systemic lupus erythematosus: a prospective study. *Ann Rheum Dis.* 1994;53:235-9.
112. Swaak AJ, Hintzen RQ, Huysen V, van den Brink HG, Smeenk JT. Serum levels of soluble forms of T cell activation antigens CD27 and CD25 in systemic lupus erythematosus in relation with lymphocytes count and disease course. *Clin Rheumatol.* 1995;14:293-300.
113. Davas EM, Tsirogianni A, Kappou I, Karamitsos D, Economidou I, Dantis PC. Serum IL-6, TNFalpha, p55 srTNFalpha, p75srTNFalpha, srIL-2alpha levels and disease activity in systemic lupus erythematosus. *Clin Rheumatol.* 1999;18:17-22.

114. El-Shafey EM, El-Nagar GF, El-Bendary AS, Sabry AA, Selim AG. Serum soluble interleukin-2 receptor alpha in systemic lupus erythematosus. *Iran J Kidney Dis.* 2008;2:80-5.
115. Zhang RJ, Zhang X, Chen J, et al. Serum soluble CD25 as a risk factor of renal impairment in systemic lupus erythematosus - a prospective cohort study. *Lupus.* 2018;27:1100-6.
116. Petty RE, Southwood TR, Baum J, et al. Revision of the proposed classification criteria for juvenile idiopathic arthritis: Durban, 1997. *J Rheumatol.* 1998;25:1991-4.
117. Petty RE, Southwood TR, Manners P, et al. International League of Associations for Rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. *J Rheumatol.* 2004;31:390-2.
118. Martini A, Ravelli A, Avcin T, et al. Toward New Classification Criteria for Juvenile Idiopathic Arthritis: First Steps, Pediatric Rheumatology International Trials Organization International Consensus. *J Rheumatol.* 2019;46:190-7.
119. Prahalad S, Martins TB, Tebo AE, et al. Elevated serum levels of soluble CD154 in children with juvenile idiopathic arthritis. *Pediatr Rheumatol Online J.* 2008;6:8.
120. Shahin AA, Shaker OG, Kamal N, Hafez HA, Gaber W, Shahin HA. Circulating interleukin-6, soluble interleukin-2 receptors, tumor necrosis factor alpha, and interleukin-10 levels in juvenile chronic arthritis: correlations with soft tissue vascularity assessed by power Doppler sonography. *Rheumatol Int.* 2002;22:84-8.
121. Silverman ED, Laxer RM, Nelson DL, Rubin LA. Soluble interleukin-2 receptor in juvenile rheumatoid arthritis. *J Rheumatol.* 1991;18:1398-402.
122. Bojko JJ, Omelczenko LI, Czernyszow WP. Predictors of juvenile idiopathic arthritis course. *Reumatologia.* 2015;53:119-24.
123. Ravelli A, Minoia F, Davi S, et al. 2016 Classification Criteria for Macrophage Activation Syndrome Complicating Systemic Juvenile Idiopathic Arthritis: A European League Against Rheumatism/American College of Rheumatology/Paediatric Rheumatology International Trials Organisation Collaborative Initiative. *Arthritis Rheumatol.* 2016;68:566-76.
124. Bleesing J, Prada A, Siegel DM, et al. The diagnostic significance of soluble CD163 and soluble interleukin-2 receptor alpha-chain in macrophage activation syndrome and untreated new-onset systemic juvenile idiopathic arthritis. *Arthritis Rheum.* 2007;56:965-71.
125. Behrens EM, Beukelman T, Paessler M, Cron RQ. Occult macrophage activation syndrome in patients with systemic juvenile idiopathic arthritis. *J Rheumatol.* 2007;34:1133-8.
126. Feist E, Mitrovic S, Fautrel B. Mechanisms, biomarkers and targets for adult-onset Still's disease. *Nat Rev Rheumatol.* 2018;14:603-18.
127. Gerfaud-Valentin M, Jamilloux Y, Iwaz J, Seve P. Adult-onset Still's disease. *Autoimmun Rev.* 2014;13:708-22.
128. Nirmala N, Brachat A, Feist E, et al. Gene-expression analysis of adult-onset Still's disease and systemic juvenile idiopathic arthritis is consistent with a continuum of a single disease entity. *Pediatr Rheumatol Online J.* 2015;13:50.
129. Lee SJ, Cho YN, Kim TJ, et al. Natural killer T cell deficiency in active adult-onset Still's Disease: correlation of deficiency of natural killer T cells with dysfunction of natural killer cells. *Arthritis Rheum.* 2012;64:2868-77.
130. Park JH, Kim HS, Lee JS, et al. Natural killer cell cytolytic function in Korean patients with adult-onset Still's disease. *J Rheumatol.* 2012;39:2000-7.
131. Chen DY, Lan JL, Lin FJ, Hsieh TY, Wen MC. Predominance of Th1 cytokine in peripheral blood and pathological tissues of patients with active untreated adult onset Still's disease. *Ann Rheum Dis.* 2004;63:1300-6.
132. Chen DY, Chen YM, Lan JL, Lin CC, Chen HH, Hsieh CW. Potential role of Th17 cells in the pathogenesis of adult-onset Still's disease. *Rheumatology (Oxford).* 2010;49:2305-12.
133. Chen DY, Lan JL, Lin FJ, Hsieh TY. Proinflammatory cytokine profiles in sera and pathological tissues of patients with active untreated adult onset Still's disease. *J Rheumatol.* 2004;31:2189-98.
134. Choi JH, Suh CH, Lee YM, et al. Serum cytokine profiles in patients with adult onset Still's disease. *J Rheumatol.* 2003;30:2422-7.
135. Fujii T, Nojima T, Yasuoka H, et al. Cytokine and immunogenetic profiles in Japanese patients with adult Still's disease. Association with chronic articular disease. *Rheumatology (Oxford).* 2001;40:1398-404.
136. Girard C, Rech J, Brown M, et al. Elevated serum levels of free interleukin-18 in adult-onset Still's disease. *Rheumatology (Oxford).* 2016;55:2237-47.
137. Schwarz-Eywill M, Heilig B, Bauer H, Breitbart A, Pezzutto A. Evaluation of serum ferritin as a marker for adult Still's disease activity. *Ann Rheum Dis.* 1992;51:683-5.
138. Chen DY, Chen YM, Chen HH, Hsieh CW, Lin CC, Lan JL. The associations of circulating CD4+CD25high regulatory T cells and TGF-beta with disease activity and clinical course in patients with adult-onset Still's disease. *Connect Tissue Res.* 2010;51:370-7.
139. Jennette JC, Falk RJ, Bacon PA, et al. 2012 revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides. *Arthritis Rheum.* 2013;65:1-11.
140. Bossuyt X, Cohen Tervaert JW, Arimura Y, et al. Position paper: Revised 2017 international consensus on testing of ANCA in granulomatosis with polyangiitis and microscopic polyangiitis. *Nat Rev Rheumatol.* 2017;13:683-92.
141. Arranz O, Ara J, Rodriguez R, Saurina A, Mirapeix E, Darnell A. Serum levels of soluble interleukin-2 receptor in patients with ANCA-associated vasculitis. *J Nephrol.* 2000;13:59-64.
142. Sanders JS, Huitma MG, Kallenberg CG, Stegeman CA. Plasma levels of soluble interleukin 2 receptor, soluble CD30, interleukin 10 and B cell activator of the tumour necrosis factor family during follow-up in vasculitis associated with proteinase 3-antineutrophil cytoplasmic antibodies: associations with disease activity and relapse. *Ann Rheum Dis.* 2006;65:1484-9.
143. Schmitt WH, Heesen C, Csernok E, Rautmann A, Gross WL. Elevated serum levels of soluble interleukin-2 receptor in patients with Wegener's granulomatosis. Association with disease activity. *Arthritis Rheum.* 1992;35:1088-96.
144. Stegeman CA, Tervaert JW, Huitema MG, Kallenberg CG. Serum markers of T cell activation in relapses of Wegener's granulomatosis. *Clin Exp Immunol.* 1993;91:415-20.
145. Etzerodt A, Moestrup SK. CD163 and inflammation: biological, diagnostic, and therapeutic aspects. *Antioxid Redox Signal.* 2013;18:2352-63.
146. Zhi Y, Gao P, Xin X, et al. Clinical significance of sCD163 and its possible role in asthma (Review). *Mol Med Rep.* 2017;15:2931-9.
147. O'Reilly VP, Wong L, Kennedy C, et al. Urinary Soluble CD163 in Active Renal Vasculitis. *J Am Soc Nephrol.* 2016;27:2906-16.
148. Dekkema GJ, Abdulhad WH, Bijma T, et al. Urinary and serum soluble CD25 complements urinary soluble CD163 to detect active renal anti-neutrophil cytoplasmic autoantibody-associated vasculitis: a cohort study. *Nephrol Dial Transplant.* 2019;34:234-42.
149. Kamisawa T, Zen Y, Pillai S, Stone JH. IgG4-related disease. *Lancet.* 2015;385:1460-71.
150. Kamisawa T, Shimosegawa T, Okazaki K, et al. Standard steroid treatment for autoimmune pancreatitis. *Gut.* 2009;58:1504-7.
151. Khosroshahi A, Carruthers MN, Deshpande V, Unizony S, Bloch DB, Stone JH. Rituximab for the treatment of IgG4-related disease: lessons from 10 consecutive patients. *Medicine (Baltimore).* 2012;91:57-66.
152. Heeringa JJ, Karim AF, van Laar JAM, et al. Expansion of blood IgG4(+) B, TH2, and regulatory T cells in patients with IgG4-related disease. *J Allergy Clin Immunol.* 2018;141:1831-43 e10.
153. Karim AF, Heeringa JJ, van Laar JAM, et al. Reply. *J Allergy Clin Immunol.* 2018;141:1958-60 e4.
154. Akiyama M, Sasaki T, Kaneko Y, et al. Serum soluble interleukin-2 receptor is a useful biomarker for disease activity but not for differential diagnosis in IgG4-related disease and primary Sjogren's syndrome adults from a defined population. *Clin Exp Rheumatol.* 2018;36 Suppl 112:157-64.
155. Handa T, Matsui S, Yoshifuji H, et al. Serum soluble interleukin-2 receptor as a biomarker in immunoglobulin G4-related disease. *Mod Rheumatol.* 2018;28:838-44.
156. Hannani D, Vetzizou M, Enot D, et al. Anticancer immunotherapy by CTLA-4 blockade: obligatory contribution of IL-2 receptors and negative prognostic impact of soluble CD25. *Cell Res.* 2015;25:208-24.
157. Anderson R, Rapoport BL. Immune Dysregulation in Cancer Patients Undergoing Immune Checkpoint Inhibitor Treatment and Potential Predictive Strategies for Future Clinical Practice. *Front Oncol.* 2018;8:80.