Reagent strips are efficient to rule out spontaneous bacterial peritonitis in cirrhotics

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ABSTRACT

Background: The gold standard to diagnose spontaneous bacterial peritonitis (SBP) is a polymorphonuclear neutrophil count ≥ 250 cells/µl in ascitic fluid. This test is laborious and expensive. Urine reagent strips measuring leukocyte esterase activity have been proposed as a rapid and inexpensive alternative. The aim of this study was to assess the diagnostic accuracy of the Combur reagent strip for diagnosing SBP. Furthermore the possible advantage of a photospectrometer reading over visual reading of the strip was investigated.

Methods: This prospective study includes all ascitic fluid samples of cirrhotic patients undergoing diagnostic or therapeutic paracentesis over a 12-month period. The samples were collected for the standard diagnostic work-up and in addition tested with a bedside Combur reagent strip. The strip was read visually and with an automated spectrometer.

Results: A total of 157 samples were obtained from 53 patients, and spontaneous bacterial peritonitis was diagnosed in 12 patients based on the ascitic polymorphonuclear neutrophil count. The sensitivity, specificity, positive predictive value and negative predictive value of the reagent strip according to the photospectrometer were 100%, 93%, 55% and 100% respectively, and 75%, 99%, 82% and 98%, respectively, for visual interpretation. The diagnostic accuracy of the photospectrometer was found to be higher than visual interpretation (p = 0.007).

Conclusion: The diagnostic accuracy of leukocyte esterase reagent strips read out by a photospectrometer was comparable with the gold standard test and was excellent for excluding SBP. Our results support implementation of reagent strips in the diagnostic work-up of ascitic fluid.

KEYWORDS

Ascites, leukocyte count, liver cirrhosis, peritonitis, reagent strips

INTRODUCTION

Spontaneous bacterial peritonitis (SBP) is a life-threatening complication in cirrhotic patients with ascites.1 Late or misdiagnosed SBP can lead to increased mortality due to consequences such as gastrointestinal bleeding, development of hepatorenal syndrome and progressive liver failure. Therefore, the threshold for performing diagnostic paracentesis and ascitic analysis should be low.6 The reported prevalence of SBP in cirrhotic patients differs from 0-2.8% in outpatients to 10-30% in hospitalised patients.3-9 The gold standard test to diagnose SBP is a polymorphonuclear neutrophil count of ≥ 250/µl in ascites using a manual counting chamber, regardless of the outcome of the culture of ascitic fluid.4 This analysis is laborious, time-consuming and expensive. Automated cell counting has been proposed to be a reasonable alternative with a high diagnostic accuracy.10

In the past two decades several studies have examined the use of leukocyte esterase reagent strips for the bedside diagnosis of SBP.8,14-31 These strips are widely used for rapid urinary analysis and the principle is based on the detection of leukocyte esterase activity of granulocytes. Varying levels of diagnostic accuracy to diagnose SBP with reagent strips have been reported, with a sensitivity ranging from 45-100%, a specificity from 90-100%, a positive predictive value from 42-100% and a negative predictive value from 93-100%.8,14-31 These inconsistent results could be related to variability in reagent strips, patient populations, different cut-off values and the
subjective interpretation of the reagent strip result. However, the consistent high negative predictive value could make the reagent strips a very useful rule-out tool.

This study was performed to 1) assess the diagnostic accuracy of reagent strips in comparison with the current gold standard test for diagnosing SBP in a mixed population of low-risk and high-risk patients, and 2) investigate the possible advantage of automated analysis of the reagent strips over visual non-automated reading.

MATERIALS AND METHODS

Study design
This prospective cohort study was carried out at the Department of Gastroenterology and Hepatology in a referral centre for liver disease in the Netherlands. The study was designed and carried out in accordance with the principles of the Helsinki Declaration and approval was given by the local medical ethics committee of the hospital.

Patients
Consecutive patients with cirrhosis undergoing diagnostic or therapeutic paracentesis were prospectively enrolled from July 2006 up to and including July 2007. The total study population was subdivided into a low-risk and high-risk population for the development of SBP. The low-risk population was defined as patients undergoing therapeutic, large volume paracentesis or outpatients undergoing diagnostic paracentesis. The high-risk population was defined as hospitalised patients undergoing a standard diagnostic paracentesis at admission or because of clinical deterioration. Patients with ascites secondary to causes other than liver disease were excluded.

Methods
Paracentesis was performed under strict sterile conditions. Ascitic fluid was routinely analysed in the central clinical laboratory with automated determination of the white blood cell count with differential. Ten millilitres of fluid was inoculated at the bedside in aerobic and anaerobic blood culture bottles (Bactec®). Fluid was collected in a sterile tube and assessed by two leukocyte esterase reagent strips (Combur® strips, Roche Diagnostics). Both strips were read out after 60 seconds, one strip visually and one with a photospectrometer (Urisys 1100®, Roche Diagnostics). The observer was unaware of the results of the spectrometer. The observer could differentiate between four different colour shades corresponding to 0, 25, 100 or 500 leukocytes/µl.

Statistical analysis
Data analysis was performed using IBM SPSS Statistics for Windows, Version 21.0 (Armonk, NY: IBM Corp.). A mean and standard deviation was computed for continuous variables and compared with the Student’s t-tests if normally distributed. A two-sided p-value < 0.05 was considered significant. Sensitivity, specificity, positive and negative predictive values with confidence intervals of 95% were calculated. Receiver operating characteristic (ROC) curves were computed and the optimal categorical cut-off point was analysed. Diagnostic performance between photospectrometer reading and visual interpretation was statistically compared using a McNemar test.

RESULTS

A total of 157 ascitic fluid samples were collected from 52 patients (range 1-14 samples per patient): 87 samples (55%) were obtained in the low-risk population and 70 (45%) in the high-risk population (table 1). The prevalence of SBP according to analysis of the polymorphonuclear neutrophil count was 4 (4.5%) in the low-risk group and 8 (11.6%) in the high-risk group (figure 1).

In the low-risk population, one culture (25%) was positive, identifying an Enterococcus faecium, whereas three cultures (37.5%) were positive in the high-risk population, identifying Enterococcus coli, Haemophilus parainfluenzae and Pseudomonas aeruginosa in one case each.

Photospectrometer versus visual reading

Of the total of 12 (25%) cases of SBP, three were not detected by optical reading of the strip but correctly diagnosed with the photospectrometer.

With visual reading, the sensitivity for diagnosing SBP was 75% (95% CI 41-93), the specificity 99% (95% CI 95-100), the positive predictive value 82% (95% CI 48-97) and the negative predictive value 98% (95% CI 94-100). The diagnostic accuracy for automated reading was slightly superior (p = 0.007 McNemar test): sensitivity 100% (95% CI 70-100), specificity 93% (95% CI 87-97), positive predictive value 55% (95% CI 37-75) and negative predictive value 100% (95% CI 97-100) (table 2). ROC curve analysis indicated that the diagnostic accuracy of the strips was optimal at a cut-off of 100 leukocytes/µl.

Low- and high-risk group analysis

The diagnostic performance of the strip with automated reading in the low- and high-risk populations was similar: the negative predictive value was 100% (95% CI 92-100%) and the specificity was 93% (95% CI 81-98%).

DISCUSSION

The results of the present study support the diagnosing value of leukocyte esterase reagent strips in ascitic fluid
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Analysis in patients with cirrhosis. In particular, this simple, quick and inexpensive method could reliably rule out SBP, with a 100% negative predictive value in populations at low- and high-risk for SBP. Automated reading of the reagent strip was superior to visual interpreting and prevented false-negative results.

The diagnostic accuracy of the Combur® strip in ascitic fluid analysis has been studied previously by several groups. The results of these studies were comparable with our results in terms of a high negative predictive value of reagent strip testing. The cumulative data suggest that the sensitivity of strips for diagnosing SBP is variable and may not be optimal. A negative test result, however, strongly predicts absence of SBP. Thus, in patients undergoing diagnostic paracentesis, a negative reagent strip result may imply that further diagnostic studies – polymorphonuclear neutrophil count and bacterial cultures – are not useful and can be omitted. Obviously, preventing unnecessary diagnostic studies in a substantial proportion of patients presenting with ascites may lead to a marked reduction in costs.

One of the limitations of our study may be that the reagent strips we used are not specifically designed for ascitic fluid analysis. Our results suggest that automated reading is superior and may be the preferred method in clinical practice. Additional studies would be useful to confirm this finding.

Table 1. Baseline characteristics in 52 patients in the low-risk and high-risk group

<table>
<thead>
<tr>
<th></th>
<th>All (n = 52)</th>
<th>Low-risk group (n = 20)</th>
<th>High-risk group (n = 32)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male (%)</td>
<td>35 (67%)</td>
<td>17 (85%)</td>
<td>18 (56%)</td>
<td>0.038</td>
</tr>
<tr>
<td>Age*, years</td>
<td>51 ± 10</td>
<td>51 ± 8</td>
<td>51 ± 11</td>
<td>0.381</td>
</tr>
<tr>
<td>Child-Pugh score*</td>
<td>10 ± 1.5</td>
<td>9 ± 1.4</td>
<td>10 ± 1.6</td>
<td>0.962</td>
</tr>
<tr>
<td>Aetiology of liver cirrhosis (%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alcohol</td>
<td>18 (35%)</td>
<td>11 (55%)</td>
<td>7 (22%)</td>
<td>0.325</td>
</tr>
<tr>
<td>Cryptogenic</td>
<td>10 (19%)</td>
<td>3 (15%)</td>
<td>7 (22%)</td>
<td></td>
</tr>
<tr>
<td>Viral</td>
<td>7 (13%)</td>
<td>2 (10%)</td>
<td>5 (16%)</td>
<td></td>
</tr>
<tr>
<td>Viral + alcohol</td>
<td>3 (6%)</td>
<td>0 (0%)</td>
<td>3 (9%)</td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>14 (27%)</td>
<td>4 (20%)</td>
<td>10 (31%)</td>
<td></td>
</tr>
</tbody>
</table>

*Mean ± standard deviation.

Table 2. Diagnostic accuracy of visual and automated reading of the leukocyte esterase reagent strip compared with the gold standard

<table>
<thead>
<tr>
<th></th>
<th>Visual reading</th>
<th>Photospectrometer reading</th>
</tr>
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<tbody>
<tr>
<td>Sensitivity</td>
<td>75% (95% CI 43-93%)</td>
<td>100% (95% CI 70-100%)</td>
</tr>
<tr>
<td>Specificity</td>
<td>99% (95% CI 95-100%)</td>
<td>93% (95% CI 87-96%)</td>
</tr>
<tr>
<td>Positive predictive value</td>
<td>82% (95% CI 48-97%)</td>
<td>55% (95% CI 33-75%)</td>
</tr>
<tr>
<td>Negative predictive value</td>
<td>98% (95% CI 94-99%)</td>
<td>100% (95% CI 97-100%)</td>
</tr>
</tbody>
</table>

Figure 1. Flowchart study participants and sample collection
Table 3. Overview of studies assessing the diagnostic value of Combur leukocyte esterase reagent strips for diagnosing SBP

<table>
<thead>
<tr>
<th>Author, year</th>
<th>Samples</th>
<th>Prevalence SBP (%)</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>Positive predictive value</th>
<th>Negative predictive value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thevenot, 2004&lt;sup&gt;10&lt;/sup&gt;</td>
<td>100</td>
<td>9 (9%)</td>
<td>89</td>
<td>100</td>
<td>100</td>
<td>99</td>
</tr>
<tr>
<td>Sarwar, 2005&lt;sup&gt;11&lt;/sup&gt;</td>
<td>214</td>
<td>38 (18%)</td>
<td>83</td>
<td>83</td>
<td>42</td>
<td>97</td>
</tr>
<tr>
<td>Braga, 2006&lt;sup&gt;12&lt;/sup&gt;</td>
<td>100</td>
<td>9 (9%)</td>
<td>100</td>
<td>98.9</td>
<td>92.3</td>
<td>100</td>
</tr>
<tr>
<td>Campillo, 2006&lt;sup&gt;13&lt;/sup&gt;</td>
<td>443</td>
<td>33 (%)</td>
<td>63</td>
<td>99.2</td>
<td>91</td>
<td>92.9</td>
</tr>
<tr>
<td>Rerknimitr, 2006&lt;sup&gt;14&lt;/sup&gt;</td>
<td>200</td>
<td>42 (21%)</td>
<td>88</td>
<td>81</td>
<td>55</td>
<td>96</td>
</tr>
<tr>
<td>Rerknimitr, 2010&lt;sup&gt;15&lt;/sup&gt;</td>
<td>350</td>
<td>30 (12%)</td>
<td>90</td>
<td>93.2</td>
<td>64.3</td>
<td>98.6</td>
</tr>
<tr>
<td>Present study, 2015</td>
<td>157</td>
<td>12 (8%)</td>
<td>100</td>
<td>93</td>
<td>55</td>
<td>100</td>
</tr>
</tbody>
</table>

References


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