The gut microbiota in internal medicine: implications for health and disease

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ABSTRACT

The human gut microbiota may be viewed as an organ, executing numerous functions in metabolism, development of the immune system and host defence against pathogens. It may therefore be involved in the development of a range of diseases such as gastrointestinal infections, inflammatory bowel disease, allergy and diabetes mellitus. Reversely, certain therapies that are often used, such as antibiotics and chemotherapy, may negatively affect the composition and function of the gut microbiota and thereby the wellbeing of patients. As the microbiota research field is currently moving from association studies to intervention studies and even clinical trials, implementation of this new knowledge into clinical practice is coming near. Several therapeutic interventions that target the gut microbiota are being evaluated, ranging from supplementation of food components to transplantation of faecal microbiota. In this review we provide an overview of current literature on the gut microbiota in both a healthy state and a range of diseases that are relevant for internal medicine. In anticipation of gut microbiota-targeted therapies, it is important to realise the key function of the gut microbiota in physiological processes and the collateral damage that may be caused when disrupting this ecosystem within us.

KEYWORDS

Antibiotics, diabetes, gut microbiota, inflammatory bowel disease, internal medicine

INTRODUCTION

The field of microbiota research has exploded in recent years. In the first six months of 2014, over 100 peer-reviewed articles were published on this topic on each consecutive day. The awareness that commensal microorganisms are not simple bystanders in our bodies, but instead play key roles in physiology and pathology, has excited scientists and clinicians in almost every discipline of medicine.¹⁻³ The unprecedented success of faecal transplantation as a potential cure for recurrent *Clostridium difficile* infection has become the gem of the microbiota research field;⁴ still, the gut microbiota is suspected of having an important role in a whole range of diseases.⁵

'The last human organ' plays a major role in the development of the immune system, the defence against pathogens and the metabolism of fatty acids, glucose and bile acids (*figure 1*).⁶ Of special interest is the degradation of otherwise non-fermentable dietary fibre such as resistant starch into short-chain fatty acids (SCFA), mainly by bacteria from the Bacteroidetes phylum.⁷ These SCFA – particularly acetic, propionic and butyric acid – have anti-inflammatory and immune-signalling properties and are an energy source for epithelial cells.⁸

Our gut contains ten times more bacteria than our bodies contain cells of our own.⁹ Moreover, the collective microbiome is 150 times larger than the human genome, indicating the astonishing number of processes that the intestinal microbes are involved in.⁹ It remains to be determined, however, if this new wealth of knowledge will affect clinical medical practice. In this review we provide an overview of the current literature on the role of the gut microbiota in health and disease from the perspective of the internist.

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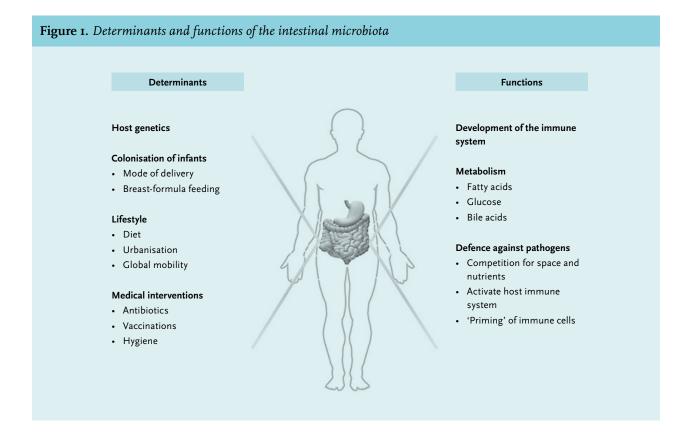
COMPOSITION OF THE GUT MICROBIOTA

Determining the exact composition of the gut microbiota has been difficult due to the limited success of culturing (often anaerobic) bacteria. With the development of specialised microarrays and high-throughput sequencing techniques, our knowledge on this subject has started to expand quickly (textbox).9.10 The new insights gathered with these new techniques start right at birth: we now know that we are not born sterile, as low levels of bacteria are detected in meconium and umbilical cord blood.¹¹ Major colonisation, however, starts after birth and is influenced by many factors such as the genetics of the host, mode of delivery, breast or formula feeding, nutrition and antibiotics. The diversity of the gut microbiota increases in the first years of life,¹¹ after which the core composition appears to remain relatively constant during adult life:12,13 60% of all gut microbiota strains in healthy adults remain detectable over a period of five years, following a power law that suggests that this core is present for a much longer period.13,14

Each individual has their own unique microbiota. Even between healthy persons the composition may vary strongly as was shown in over 1000 adults.^{10,15} However, when looking at a functional level, most people carry equal amounts of bacterial genes involved in metabolic pathways. This suggests that the microbiota as a functional organ is similar among human beings.¹⁰ Diet is one of the most important determinants of the microbial composition.¹⁶ A study in healthy adults showed that an extremely fat-rich diet is capable of changing the microbiota in just a few days.¹⁷ Still, pinpointing causal relationships between the absence or overabundance of bacterial strains and clinical observations is difficult. Few prospective and intervention studies have been performed hitherto.^{13,17,18} With older age, inter-individual differences in gut microbiota composition become larger. For instance, long-term care residency is associated with lower bacterial diversity.¹⁹ Still, the largest and most direct effects on microbial composition are achieved by antibiotics.

INFLUENCE OF ANTIBIOTICS

A simple antibiotic course, prescribed to kill just one or a few pathogens, will quickly result in major disturbances of the gut microbiota (*table 1*).²⁰ Recovery and stabilisation of its diversity level may take months, and the new composition of bacteria may significantly differ from pre-treatment. Not only bacteria that fall within the spectrum of the antibiotic are affected; species that are dependent on the ones being killed may disappear as well. Vancomycin, for example, does not only lead to the disappearance of Gram-positive species, but also Gram-negative species and results in more dramatic shifts



Investigating the gut microbiota

Techniques

Classic microbiological culturing techniques have proved inadequate in characterising the gut microbiota as a whole; sequencing techniques are therefore essential. High throughput methods have been introduced, sequencing either total DNA ('shotgun metagenomics') or 16S ribosomal RNA- genes.^{9,10} 16S rRNA contains nine sequences that are highly variable between species and have been well conserved during evolution; therefore it is suitable as a key identifier of bacteria.

Two major reference databases serve as the basis of our knowledge of the structure and function of the human gut microbiota. The US Human Microbiome Project (HMP; www.hmpdacc.org) is a consortium that has reported the structure and function of the human microbiome in 242 healthy individuals at 18 body sites from a single time point, using 16S ribosomal RNA gene pyrosequencing.¹⁰ Similarly, the European Metagenomics of the Human Intestinal Tract (MetaHIT; www.metahit.eu) project has studied faecal samples from 124 healthy, overweight and obese Europeans, as well as IBD patients, by total DNA metagenomic sequencing.⁹

Analysis

One of the major outcome measurements is species diversity, which is defined as the effective number of different species that are represented in a dataset. Results are often expressed as:

than amoxicillin.^{21,22} Antibiotics may thus have negative effects on the health of patients, for example on their metabolic state. Use of antibiotics has been associated with the development of obesity, diabetes mellitus and asthma.²

THE ROLE OF THE GUT MICROBIOTA IN DISEASE

Gastrointestinal infections

A healthy gut microbiota protects the host directly against pathogens such as *C. difficile*, by competition for nutrients, space and binding spots on the epithelium and production of bacteriocins, but also indirectly, by activation of the host immune system resulting in release of IgA, cytokines and antimicrobial peptides.²³ Depletion of the microbiota by antibiotics therefore creates an opportunity for *C. difficile* to proliferate.

Multi-drug resistant organisms are a rapidly increasing problem around the world. Vancomycin-resistant enterococci (VRE) are now a leading cause of bloodstream infections

- Species richness and species evenness, which together make up diversity (also termed alpha diversity – see below)
 - Species richness: number of different species represented in an ecological community
 - Species evenness: the relative abundance with which each species is represented in the community. An ecosystem where all the species are represented by the same number of individuals has high species evenness
- 2. Alpha and beta diversity
 - Alpha diversity : within-sample species diversity
 - Beta diversity: similarity between samples (from one individual at different time points or between samples from different subjects)
- 3. Different indexes, such as:
 - Shannon index: quantifies the degree of uncertainty when predicting an individual's type within a particular dataset
 - Simpson index: quantifies the probability that two entities taken at random from the dataset represent the same type

in haematopoietic stem cell transplant recipients.²⁴ In both mice and men, domination by VRE (being at least 30% of the microbiota) correlates with subsequent development of a bloodstream infection²⁵ and in mice can be reversed by faecal microbiota transplantation (FMT).²⁴ This suggests that faecal transplantation therapy might be effective in both prevention and treatment of VRE infections. Assessment of faecal microbiota may be used to identify those patients at highest risk for bloodstream infection.²⁵

The first randomised clinical trial for FMT in recurrent *C. difficile* infection was stopped prematurely because of a clear advantage in the FMT arm: 15 of 16 patients had complete resolution of disease, compared with four of 13 patients in the vancomycin arm and three of 13 patients in the vancomycin + bowel lavage arm.⁴ European recommendations now include FMT for patients with multiple relapses of *C. difficile* infection.²⁶

Inflammatory bowel disease

Inflammatory bowel disease (IBD) is hypothesised to be either an aberrant immune response against normal

Author	Antibiotic regimen	Subjects	Short-term results (days, weeks)	Long-term results (months, years)
De la Cochetiere et al., 2005 ⁵⁶	Amoxicillin, orally, 5 days	6 adult healthy volunteers	Major shift in dominant species after 24 hours Average similarity (compared with pre-treatment) 74% after 4 days	1-2 month: average similarity 88-89%
Penders et al., 2006 ⁵⁷	Mostly amoxicillin	28 paediatric patients (1 month old)	Decreased counts of <i>Bifidobacteria</i> and <i>B. fragilis</i> species compared with non-treated children	None reported
Jernberg et al., 2007 ⁵⁸	Clindamycin, orally, 7 days	4 adult healthy volunteers (vs. 4 controls)	Day 7, 21: large and persistent shift in composition	3, 6, 9, 12, 18 and 24 months: large and persistent shift in composition
Dethlefsen et al., 2008 ⁵⁹	Ciprofloxacin, orally, 5 days	3 adult healthy volunteers	Abundance of about a third of the bacterial taxa in the gut affected; decreased taxonomic richness, diversity, and evenness of the community	I and 6 months: richness, diversity and evenness comparable to pre- antibiotic state, some long-term losses
Dethlefsen et al., 2010 ²⁰	Ciprofloxacin, orally, 5 days; after 6 months again 5 days	3 adult healthy volunteers	Loss of diversity and shift in composition within 3–4 days; 7 days after end of a course, communities start returning to initial state; often incomplete	10 months: composition stabilised but altered; long-term losses of some taxa
Jakobsson et al., 2010 ⁶⁰	Metronidazole + clarithromycin, orally, 7 days	3 adult healthy volunteers (vs. 3 controls)	Dramatic decline in diversity, especially loss of Actinobacteria, in both throat and faeces	ı year: diversity levels recovered to pre-treatment states
Fouhy et al., 2012 ⁶¹	Parenteral antibiotic treatment with ampicillin and gentamicin (within 48 h of birth)	9 paediatric patients (plus 9 untreated infants)	4 weeks after treatment, antibiotic- treated infants had higher proportions of Proteobacteria and lower proportions of Actinobacteria as well as the genus <i>Lactobacillus</i>	2 months: Proteobacteria levels remained higher, but Actinobacteria and <i>Lactobacillus</i> levels had recovered
Panda et al., 2014 ⁶²	Different broad- spectrum antibiotic regimens, orally	21 adult patients	Fluoroquinolones and b-lactams decreased microbial diversity by 25% and reduced the number of taxa from 29 to 12; Increase in proportion Bacteroidetes taxa B-lactams increased the average microbial load two-fold	None reported

Table I.	Overview of	of studies on	the effect o	f antibiotics on the	human intestinal microbiota

commensal bacteria, or a normal immune response against abnormal gut microbiota, both driven by an autoimmune genetic background.²⁷ Indeed, IBD patients have a significantly lower microbial diversity compared with healthy controls²⁸ as well as a lower abundance of immune-modulating bacteria such as *Faecalibacterium prausnitzii* and *Akkermansia muciniphila.*^{5,29} *F. prausnitzii* and *A. muciniphila* produce butyrate and propionate, respectively, both SCFA that are thought to protect the mucosal barrier. A meta-analysis of 41 cases in which FMT was used for ulcerative colitis found that 63% of patients entered remission and 76% could stop medications.²⁸ bias. Thirteen clinical trials are currently ongoing to evaluate the effect of FMT in IBD (*table 2*).

Transplant immunology

Intestinal inflammation secondary to graft-versus-host disease (GVHD) is likely to be influenced by the gut microbiota. A mouse model of GVHD caused by allogeneic stem cell transplantation (SCT) showed that mice developing intestinal GVHD have a decreased bacterial diversity compared with those that do not.³⁰ A greater proportion of bacteria consisted of Lactobacillales, while Clostridiales were decreased. The same pattern was found in 31 patients receiving allogeneic SCT, probably

Table 2. Current status of microbiota-related therapies on clinicaltrials.gov and trialregister.nl (September 2014)					
Gastrointestinal infections	 First randomised trials in <i>C. difficile</i> completed, with promising results for FMT in severe recurrent infections 21 clinical trials for FMT in <i>C. difficile</i> infections registered First case report describing FMT clearing extended spectrum beta-lactamase producing <i>Escherichia coli</i> (ESBL) colonisation in end-stage renal disease patient⁶⁴ 				
Inflammatory bowel disease	• Thirteen clinical trials currently registered for FMT in IBD (both Crohn's disease and ulcerative colitis)				
Transplant immunology	No microbiota-related clinical trials being conducted yet				
Colorectal cancer	No microbiota-related clinical trials being conducted yet				
Allergic airway diseases	 > 50 trials registered for prebiotics or probiotics for treatment or prevention of allergic airway disease, mostly in infants 				
Metabolic disorders	 Two randomised trials for FMT in metabolic syndrome registered, one completed One randomised trial for FMT in recently diagnosed diabetes mellitus type 1 registered One randomised trial for FMT in diabetes mellitus type 2 registered 				
FMT = faecal microbiota t	ransplantation; IBD = inflammatory bowel disease.				

due to chemotherapy, irradiation and antibiotics. The percentage of *Enterococcus faecium* and *Enterococcus faecalis* (of the order Lactobacillales) in the microbiota was 21% in patients who did not develop intestinal GVHD, 46% in those that subsequently developed GVHD and 74% at the time of actual GVHD.³¹ The diversity level at the time of engraftment seems to predict outcome: after classifying 80 SCT patients into low, intermediate, and high diversity groups, overall three-year survival was 36%, 60%, and 67% respectively.³²

Colorectal cancer and response to chemotherapy

The intestinal microbiota play a key role in mucosal immune responses, epithelial cell homeostasis, barrier function and metabolism, all of which are important for tumour formation. For example, the microbiota produce (precursors of) carcinogenic substances, such as nitrate.5 A strictly animal-based diet (meats, eggs and cheeses) of only five days in healthy volunteers increased bile acids produced by the microbiota, including deoxycholic acid which promotes DNA damage and hepatic carcinoma.¹⁷ Accordingly, bile-tolerant bacteria such as Bilophila wadsworthia were increased, which in mice have been described to induce IBD by producing hydrogen sulphide.¹⁷ This supports the association between diets that are rich in red meat and the development of colorectal cancer.33 Faecal transplantation from colon tumour-developing mice to germ-free mice resulted in a higher incidence of colon

tumours in the latter, compared with transplantation from control mice – most likely by inducing inflammation prior to induction of tumorigenesis by dextran sodium sulphate in drinking water.³⁴

The gut microbiota also influences the effectiveness of anticancer therapy.^{35:37} In different cancer mouse models, it was shown that in the absence of a healthy microbiota, tumour-infiltrating myeloid cells are less capable of producing cytokines and inducing cytotoxicity upon immunochemotherapy or platinum chemotherapy, resulting in decreased necrosis of tumours.³⁵ Both cytotoxic drugs and irradiation may cause translocation of bacteria or bacterial particles to lymph nodes, thereby enhancing immune responses directed against malignant cells.^{36,37} Destruction of the microbiota by antibiotics and other drugs may therefore reduce the efficacy of these treatments.^{35:37}

Allergic and infectious airway diseases

Low diversity of the gut microbiota, whether caused by low exposure to environmental microbes or (prenatal or postnatal) use of antibiotics, is associated with increased risk of allergic airway disease.^{38,39} The intestinal microbiota is postulated to enhance the haematopoiesis of dendritic cell precursors in bone marrow by production of SCFAs.⁴⁰ Intriguingly, dendritic cells from mice that have received propionate exhibit an impaired ability to activate T-helper 2 cells in the lung, thus preventing allergic airway

inflammation.⁴⁰ Supplementation of dietary fibre or SCFA may therefore prevent or reverse allergic airway disease.

A similar 'priming' effect by healthy gut microbiota has been observed in murine models of pneumonia and peritonitis.⁴¹⁻⁴³ Depletion of the gut microbiota by antibiotics in neonatal mice appears to hamper neutrophil production by the bone marrow, thus leading to increased counts of pathogenic bacteria and decreased survival.⁴³ It is suggested that lipopolysaccharide from gut bacteria is the priming component, although another role for SCFA is not excluded. For now, evidence for this 'gut-lung axis' in humans is lacking.

Metabolic disorders

Obesity has been associated with a lower microbial diversity in the intestine.^{44,45} Studies have reported an association between obesity and increased levels of Firmicutes, combined with decreased Bacteroidetes (the two most dominant phyla), although this could not be replicated in all cohorts.^{5,46} Overweight and obese individuals have higher levels of SCFA, without a different intake of dietary fibre, reflecting either an increased production or decreased absorption by the gut and utilisation by bacteria.⁴⁶

In both mice and men, there is evidence that the obese microbiota is more effective at harvesting energy from food, as suggested by enriched biochemical pathways.5.47 Accordingly, transplantation of gut microbiota from obese to germ-free mice leads to more weight gain than transplantation of microbiota from control mice.47 Transplantation of faeces from lean human donors into patients with metabolic syndrome decreases peripheral insulin resistance six weeks after transplantation. This is potentially caused by increased levels of butyrateproducing bacteria (Roseburia intestinalis).48 By contrast, among 20 children of 3-5 years of age, levels of butyrateproducing bacteria were also found to be higher in those that subsequently developed type I diabetes.⁴⁹ Of interest, seven days of vancomycin therapy resulted in decreased peripheral insulin sensitivity in ten males with metabolic syndrome.22 This was associated with a decrease in Firmicutes bacteria and decreased metabolism of bile acids.

From theory to therapy

Although the role for the gut microbiota in the pathophysiology of disease may be easy to imagine, translation of this knowledge into new therapeutic strategies represents a tremendous challenge. Changing one's intake of specific nutrients would be the most natural way to prevent diseases. Consumption of dietary fibre seems one of the best solutions, since they are fermented into SCFA that may have beneficial effects. Both extreme diets¹⁷ and the use of various fibres were shown to modify the human intestinal microbiota, although the latter not in all subjects. 5°

Probiotics that include a variety of lactic acid bacteria are by now commonplace in both the supermarket and the clinic, but their effectiveness is highly debatable.⁵¹ Moreover, higher mortality in patients treated with lactic acid bacteria during severe pancreatitis compared with placebo treatment has led to increased caution regarding these bacteria, notably in a clinical setting.⁵²

The successful trial on severe *C. difficile* infection has put the spotlight on FMT.²⁸ However, we currently do not know how to select a microbiota that is best to transplant. In addition, the possibility of transplanting pathogenic microorganisms, or the possible risk of metabolic and autoimmune diseases, might be a reason for caution. So far, no serious (long-term) adverse events have been reported.

Transplanting a cocktail of bacteria (the synthetic microbiota) that can be cultured or isolated easily, may be a key objective the coming years.³³ The first steps were already taken in 1989, when ten selected intestinal strains that showed inhibitory effects against *C. difficile* cured five patients with *C. difficile* infection.^{53,54} More recently, two *C. difficile* infected patients were transplanted with 33 cultured bacterial species, isolated from healthy donor faeces – both resulting in resolution of disease and no relapse in 24 months.²³ These results encourage the development of synthetic, standardised microbiota.

CONCLUSION

The booming popularity of the gut microbiota in biomedical research seems justified; in fact, the interest in host-microbiota interactions and their potential value as a therapeutic target seems rather belated in view of our ancient commensal relationship. Obviously, results from animal studies cannot be translated directly to human health and disease. Still, some of the possible microbiotadisease interactions will prove to be relevant and applicable in everyday clinical practice. The great possibilities offered by this new field of research provide hope for new, relatively straightforward and inexpensive therapies. However, most research is currently in a preclinical phase, and causative relationships remain to be established. Moving from correlation to causality will be particularly difficult for phenotypically and/or genetically heterogeneous disorders, such as diabetes, IBD and GVHD.55 In anticipation of gut microbiota-targeted therapies, it seems important to realise the key function of the gut microbiota in physiological processes and the potential collateral damage we cause when disrupting this well-balanced ecosystem.

DISCLOSURES

The authors declare no conflicts of interest.

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REFERENCES

- Blaser M, Bork P, Fraser C, Knight R, Wang J. The microbiome explored: recent insights and future challenges. Nat Rev Microbiol. 2013;11:213-7.
- Keeney KM, Yurist-Doutsch S, Arrieta MC, Finlay BB. Effect of Antibiotics on Human Microbiota and Subsequent Disease. Annu Rev Microbiol. 2014;68.
- Schuijt TJ, van der Poll T, de Vos WM, Wiersinga WJ. The intestinal microbiota and host immune interactions in the critically ill. Trends Microbiol. 2013;21:221-9.
- van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal infusion of donor feces for recurrent Clostridium difficile. N Engl J Med. 2013;368:407-15.
- de Vos WM, de Vos EA. Role of the intestinal microbiome in health and disease: from correlation to causation. Nutr Rev. 2012;70 Suppl 1:S45-56.
- 6. Baquero F, Nombela C. The microbiome as a human organ. Clin Microbiol Inf. 2012;18 Suppl 4:2-4.
- 7. Rajilic-Stojanovic M. Function of the microbiota. Best Prac Res Clin Gastroenterol. 2013;27:5-16.
- Smith PM, Howitt MR, Panikov N, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science. 2013;341:569-73.
- Qin J, Li R, Raes J, et al. A human gut microbial gene catalogue established by metagenomic sequencing. Nature. 2010;464:59-65.
- Human Microbiome Project Consortium. Structure function and diversity of the healthy human microbiome. Nature. 2012;486:207-14.
- Cheng J, Palva AM, de Vos WM, Satokari R. Contribution of the intestinal microbiota to human health: from birth to 100 years of age. Curr Top Microbiol Immunol. 2013;358:323-46.
- Martinez I, Muller CE, Walter J. Long-term temporal analysis of the human fecal microbiota revealed a stable core of dominant bacterial species. PloS One. 2013;8(7):e69621.
- Rajilic-Stojanovic MZ, Heilig HG, Tims S, Zoetendal EG, de Vos WM. Long term monitoring of the human intestinal microbiota composition. Environ Microbiol. 2013;15:1146-59.
- 14. Faith JJ, Guruge JL, Charbonneau M, et al. The long-term stability of the human gut microbiota. Science. 2013;341:1237439.
- 15. Lahti L, Salojarvi J, Salonen A, Scheffer M, de Vos WM. Tipping elements in the human intestinal ecosystem. Nat Commun. 2014;5:4344.
- Tyakht AV, Kostryukova ES, Popenko AS, et al. Human gut microbiota community structures in urban and rural populations in Russia. Nat Commun. 2013;4:2469.
- 17. David LA, Maurice CF, Carmody RN, et al. Diet rapidly and reproducibly alters the human gut microbiome. Nature. 2014;505:559-63.
- Wu GD, Chen J, Hoffmann C, et al. Linking long-term dietary patterns with gut microbial enterotypes. Science. 2011;334:105-8.
- Claesson MJ, Jeffery IB, Conde S, et al. Gut microbiota composition correlates with diet and health in the elderly. Nature. 2012;488:178-84.
- Dethlefsen L, Relman DA. Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. Proc Natl Acad Sci U S A. 2011;108 Suppl 1:4554-61.
- Robinson CJ, Young VB. Antibiotic administration alters the community structure of the gastrointestinal micobiota. Gut Microbes. 2010;1:279-84.
- Vrieze A, Out C, Fuentes S, et al. Impact of oral vancomycin on gut microbiota, bile acid metabolism, and insulin sensitivity. J Hepatol. 2014;60:824-31.

- Buffie CG, Pamer EG. Microbiota-mediated colonization resistance against intestinal pathogens. Nat Rev Immunol. 2013;13:790-801.
- Ubeda C, Bucci V, Caballero S, et al. Intestinal microbiota containing Barnesiella species cures vancomycin-resistant Enterococcus faecium colonization. Infect Immun. 2013;81:965-73.
- Taur Y, Xavier JB, Lipuma L, et al. Intestinal domination and the risk of bacteremia in patients undergoing allogeneic hematopoietic stem cell transplantation. Clin Infect Dis. 2012;55:905-14.
- Debast SB, Bauer MP, Kuijper EJ, for the European Society of Clinical Microbiology and Infectious Diseases. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for Clostridium difficile infection. Clin Microbiol Infect. 2014;20 Suppl 2:1-26.
- 27. Shanahan F, Quigley EM. Manipulation of the microbiota for treatment of IBS and IBD-challenges and controversies. Gastroenterology. 2014;146:1554-63.
- Smits LP, Bouter KE, de Vos WM, Borody TJ, Nieuwdorp M. Therapeutic potential of fecal microbiota transplantation. Gastroenterology. 2013;145:946-53.
- Sokol H, Pigneur B, Watterlot L, et al. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. Proc Natl Acad Sci U S A. 2008;105:16731-6.
- Jenq RR, Ubeda C, Taur Y, et al. Regulation of intestinal inflammation by microbiota following allogeneic bone marrow transplantation. J Exp Med. 2012;209:903-11.
- 31. Holler E, Butzhammer P, Schmid K, et al. Metagenomic analysis of the stool microbiome in patients receiving allogeneic stem cell transplantation: loss of diversity is associated with use of systemic antibiotics and more pronounced in gastrointestinal graft-versus-host disease. Biol Blood Marrow Transplant. 2014;20:640-5.
- 32. Taur Y, Jenq RR, Perales MA, et al. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. Blood. 2014;14:1174-82.
- Albenberg LG, Wu GD. Diet and the intestinal microbiome: associations, functions, and implications for health and disease. Gastroenterology. 2014;146:1564-72.
- Zackular JP, Baxter NT, Iverson KD, et al. The gut microbiome modulates colon tumorigenesis. mBio. 2013;4:e00692-13.
- Iida N, Dzutsev A, Stewart CA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science. 2013;342(6161):967-70.
- Paulos CM, Wrzesinski C, Kaiser A, et al. Microbial translocation augments the function of adoptively transferred self/tumor-specific CD8+ T cells via TLR4 signaling. J Clin Invest. 2007;117:2197-204.
- Viaud S, Saccheri F, Mignot G, et al. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. Science. 2013;342:971-6.
- Metsala J, Lundqvist A, Virta LJ, Kaila M, Gissler M, Virtanen SM. Prenatal and postnatal exposure to antibiotics and risk of asthma in childhood. Clin Exp Allergy. 2015;45:137-45.
- 39. West CE. Gut microbiota and allergic disease: new findings. Curr Opin Clin Nutr Metabol Care. 2014;17:261-6.
- Trompette A, Gollwitzer ES, Yadava K, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. Nat Med. 2014;20:159-66.
- Chen LW, Chen PH, Hsu CM. Commensal microflora contribute to host defense against Escherichia coli pneumonia through Toll-like receptors. Shock. 2011;36:67-75.
- Clarke TB, Davis KM, Lysenko ES, Zhou AY, Yu Y, Weiser JN. Recognition of peptidoglycan from the microbiota by Nod1 enhances systemic innate immunity. Nat Med. 2010;16:228-31.
- 43. Deshmukh HS, Liu Y, Menkiti OR, et al. The microbiota regulates neutrophil homeostasis and host resistance to Escherichia coli K1 sepsis in neonatal mice. Nat Med. 2014;20:524-30.
- 44. Turnbaugh PJ, Hamady M, Yatsunenko T, et al. A core gut microbiome in obese and lean twins. Nature. 2009;457:480-4.

- 45. Le Chatelier E, Nielsen T, Qin J, et al. Richness of human gut microbiome correlates with metabolic markers. Nature. 2013;500:541-6.
- 46. Fernandes J, Su W, Rahat-Rozenbloom S, Wolever TM, Comelli EM. Adiposity, gut microbiota and faecal short chain fatty acids are linked in adult humans. Nutr Diab. 2014;4:e121.
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. Nature. 2006;444:1027-31.
- 48. Vrieze A, Van Nood E, Holleman F, et al. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. Gastroenterology. 2012;143:913-6 e7.
- 49. de Goffau MC, Fuentes S, van den Bogert B, et al. Aberrant gut microbiota composition at the onset of type 1 diabetes in young children. Diabetologia. 2014;57:1569-77.
- Salonen A, de Vos WM. Impact of diet on human intestinal microbiota and health. Annu Rev Food Sci Tech. 2014;5:239-62.
- Klaenhammer TR, Kleerebezem M, Kopp MV, Rescigno M. The impact of probiotics and prebiotics on the immune system. Nat Rev Immunol. 2012;12:728-34.
- Besselink MG, van Santvoort HC, Buskens E, et al. Probiotic prophylaxis in predicted severe acute pancreatitis: a randomised, double-blind, placebo-controlled trial. Lancet. 2008;371:651-9.
- de Vos WM. Fame and future of faecal transplantations--developing next-generation therapies with synthetic microbiomes. Microb Biotech. 2013;6:316-25.
- Tvede M, Rask-Madsen J. Bacteriotherapy for Clostridium difficile diarrhoea. Lancet. 1990;335:110.

- 55. Proceed with caution. The human gut microbiota, an attractive target for therapeutic interventions. Nat Rev Microbiol. 2014;12:149.
- De La Cochetiere MF, Durand T, Lepage P, Bourreille A, Galmiche JP, Dore J. Resilience of the dominant human fecal microbiota upon short-course antibiotic challenge. J Clin Microbiol. 2005;43:5588-92.
- Penders J, Thijs C, van den Brandt PA, et al. Gut microbiota composition and development of atopic manifestations in infancy: the KOALA Birth Cohort Study. Gut. 2007;56:661-7.
- Jernberg C, Lofmark S, Edlund C, Jansson JK. Long-term ecological impacts of antibiotic administration on the human intestinal microbiota. ISME J. 2007;1:56-66.
- Dethlefsen L, Huse S, Sogin ML, Relman DA. The pervasive effects of an antibiotic on the human gut microbiota, as revealed by deep 16S rRNA sequencing. PLoS Biol. 2008;6:e280.
- Jakobsson HE, Jernberg C, Andersson AF, Sjolund-Karlsson M, Jansson JK, Engstrand L. Short-term antibiotic treatment has differing long-term impacts on the human throat and gut microbiome. PloS One. 2010;5:e9836.
- Fouhy F, Guinane CM, Hussey S, et al. High-throughput sequencing reveals the incomplete, short-term recovery of infant gut microbiota following parenteral antibiotic treatment with ampicillin and gentamicin. Antimicrob Agents Chemother. 2012;56:5811-20.
- 62. Panda S, El Khader I, Casellas F, et al. Short-term effect of antibiotics on human gut microbiota. PloS One. 2014;9:e95476.
- Singh R, van Nood E, Nieuwdorp M, et al. Donor feces infusion for eradication of Extended Spectrum beta-Lactamase producing Escherichia coli in a patient with end stage renal disease. Clin Microbiol Infect. 2014;20:O977-8.