

Effect of the factor V Leiden mutation on the incidence and outcome of severe infection and sepsis

M. Schouten^{1,2,3}, C. van 't Veer^{1,2,3}, T. van der Poll^{1,2,3}, M. Levi^{1*}

¹Department of Medicine, ²Center for Infection and Immunity Amsterdam (CINIMA), ³Center for Experimental and Molecular Medicine, Academic Medical Center, University of Amsterdam, Amsterdam, the Netherlands, *corresponding author: tel. +31 (0)20-5662171, fax: +31 (0)20-6919658, e-mail. m.m.levi@amc.uva.nl

ABSTRACT

Activation of coagulation frequently occurs in severe infection and sepsis and may contribute to the development of multiple organ dysfunction. Factor V Leiden is a relatively common mutation resulting in a mild prohaemostatic state and consequently with an increased tendency to develop thrombosis. Hypothetically, patients with factor V Leiden may suffer from more severe coagulopathy in case of severe infection or sepsis. Aggravation of the procoagulant state in sepsis may subsequently result in more severe organ dysfunction and an increased risk of death. Here we discuss the experimental and clinical evidence regarding the relationship between the presence of a factor V Leiden mutation and the incidence and outcome of sepsis.

KEYWORDS

Factor V Leiden, thrombophilia, sepsis, infection, coagulation

INTRODUCTION

Virtually all patients with sepsis have coagulation abnormalities. These abnormalities range from subtle activation of the coagulation system that can only be detected by sensitive markers for coagulation factor activation to somewhat stronger coagulation activation detectable by a small decrease in the platelet count and subclinical prolongation of global clotting times to fulminant disseminated intravascular coagulation (DIC), which is characterised by simultaneous widespread

microvascular thrombosis and profuse bleeding from various sites.¹ Septic patients with severe forms of DIC may present with manifest thromboembolic disease or clinically less apparent microvascular fibrin deposition, which predominantly presents as multiple organ dysfunction.²⁻⁴ Clinically relevant coagulation abnormalities are present in 50 to 70% of patients with severe infection or sepsis, whereas about 35% of patients will actually meet the criteria for DIC.^{5,6} There is ample evidence that activation of coagulation in concert with inflammatory activation can result in microvascular thrombosis and thereby contributes to multiple organ failure in patients with severe sepsis.^{4,7-8} Firstly, there are several reports of post-mortem findings in septic patients with coagulation abnormalities and DIC.^{9,10} These autopsy findings include diffuse bleeding at various sites, haemorrhagic necrosis of tissue, microthrombi in small blood vessels and thrombi in mid-size and larger arteries and veins.¹¹ The demonstration of ischaemia and necrosis has been associated with fibrin deposition in small and mid-size vessels of various organs.¹² Importantly, the presence of these intravascular thrombi appears to be clearly and specifically related to the development of organ dysfunction. Secondly, experimental animal studies of DIC show fibrin deposition in various organs. Experimental bacteraemia or endotoxaemia causes intra- and extravascular fibrin deposition in kidneys, lungs, liver, brain and various other organs.¹³ Amelioration of the haemostatic defect by various interventions in these experimental models appears to improve organ failure and, in some but not all cases, mortality.¹⁴⁻¹⁷ Interestingly, some studies indicate that amelioration of systemic coagulation activation will have a profound beneficial effect on resolution of local fibrin deposition and improvement of

organ failure.^{18,19} Lastly, clinical studies support the notion of coagulation as an important determinant of clinical outcome. DIC has shown to be an independent predictor of organ failure and mortality.^{2,20} In a consecutive series of patients with severe sepsis, the mortality of patients with DIC was 43%, as compared with 27% in those without DIC. In this study, mortality was also directly related to the severity of the coagulopathy in septic patients.²¹

Apart from microvascular thrombosis and organ dysfunction, coagulation abnormalities may also have other harmful consequences. For example, thrombocytopenia in patients with sepsis confers an increased risk of bleeding.²² Indeed, in particular critically ill patients with a platelet count of $<50 \times 10^9/l$ have a four to fivefold higher risk for bleeding as compared with patients with a higher platelet count.^{23,24} A low platelet count may be both the best indicator of thrombin generation and a sign of increased platelet-vessel wall interaction.¹¹

Since the prohaemostatic state in severe infection and sepsis seems to be relevant for the pathogenesis of organ dysfunction and mortality, it may be hypothesised that even a mild pre-existent prothrombotic state in patients, such as that caused by thrombophilia for example due to a factor V Leiden mutation, would aggravate the coagulation derangement during infection and sepsis and thereby affect outcome. Interestingly, experimental and clinical studies point to an interaction between a factor V Leiden mutation and the outcome of severe infection or sepsis, although the results are sometimes conflicting. In this article, we will briefly review experimental and clinical evidence on the relationship between factor V Leiden and the outcome of severe infection and sepsis.

THROMBOPHILIA AND OUTCOME IN INFECTION AND SEPSIS

Congenital thrombophilia is usually due to a genetic variation in a gene encoding a coagulation factor or – in general clinically less relevant – a fibrinolytic protein.²⁵ Such gene polymorphisms have been described for the coagulation factors prothrombin, factor V, fibrinogen and factor XIII and for the coagulation inhibitors antithrombin, protein C and protein S. In the last-mentioned, these mutations cause a deficiency of these natural anticoagulant factors. In the fibrinolytic system the most relevant polymorphism is the 4G/5G variation in the gene encoding plasminogen activator inhibitor type I (PAI-1). This polymorphism results in mildly elevated levels of PAI-1 and is related to an increased risk of myocardial infarction and ischaemic stroke.

Anecdotal reports have indicated that the presence of congenital thrombophilia may exacerbate the coagulopathy associated with severe infection and may even result

in purpura fulminans.²⁶⁻³⁰ Indeed, various coagulation defects seem to be associated with an aggravated coagulation response to infectious agents or sepsis, although a systematic overview is missing.^{31,32} Prospective studies on the incidence or outcome of severe infections and sepsis in patients with a prothrombotic polymorphism or coagulation inhibitor deficiency are not available. However, some case-control studies have reported on the prevalence of thrombophilic abnormalities in cohorts of patients with severe sepsis. Moreover, a substantial number of animal studies have been performed. These studies have particularly focussed on deficiencies in the protein C and antithrombin pathways, the factor V Leiden mutation and genetic polymorphisms in the fibrinolytic system.

Antithrombin is the cardinal inhibitor of thrombin and factor Xa activity and, like the protein C pathway, a central regulator of coagulation activation *in vivo*. There is ample evidence that antithrombin is unable to adequately regulate these coagulation proteases in case of sepsis. Clinical studies show mean levels of antithrombin as low as 30% of normal values in patients with severe sepsis, whereas in selected individuals these levels may be even lower.^{20,33,34} Low levels of antithrombin have been shown to be associated with a higher mortality in septic patients in several prospective studies.²⁰ Restoration of antithrombin levels in experimental DIC in animals has been demonstrated to adequately block the systemic activation of coagulation and in these studies was also associated with improved outcome in terms of less organ failure and a reduction in mortality.^{15,35} In mice a heterozygous deficiency of antithrombin endotoxaemia leads to much more deposition of fibrin in various organs, including the kidneys, liver and heart, as compared with endotoxaemic wild-type mice.³⁶ There are no clinical data that point to a role of antithrombin deficiency in the outcome of sepsis or severe infection in humans.

There are several indications that the protein C system plays an important role in sepsis and also that defects in the protein C system may influence the outcome in sepsis. An impaired function of the protein C system is directly related to the severity and outcome of sepsis.³⁷ The most compelling evidence comes from experimental studies showing that administration of activated protein C to septic animals resulted in amelioration of DIC and an improved survival.¹⁷ Clinical studies confirm the beneficial effect of activated protein C in sepsis.³⁸ Severe (congenital) protein C deficiency in mice results in thrombophilia as well as a proinflammatory phenotype with higher total white blood cell counts and higher basal IL-6 levels as compared with wild-type mice.³⁹ Further protein C deficiency was shown to affect endotoxaemia in a mouse model. In these experiments mice with a one-allele targeted deletion of the protein C, resulting in heterozygous protein C

deficiency,⁴⁰ were subjected to endotoxaemia.⁴¹ Mice with a heterozygous deficiency of protein C had more severe DIC, as evidenced by a greater decrease in fibrinogen level and a larger reduction in platelet count. Also thrombin-antithrombin complex levels were 3.4-fold higher in protein C^{+/-} mice as compared with wild-type mice and histological examination showed more fibrin deposition in lungs and kidneys in these mice. Survival at 12 hours after the endotoxin injection was diminished in the protein C^{+/-} group. Interestingly, protein C^{+/-} mice had significantly higher levels of the pro-inflammatory cytokines TNF- α , IL-6 and IL-1 β , indicating an interaction between the protein C system and the inflammatory response. This last observation is consistent with many other studies indicating cross-talk between effects of protein C on coagulation and inflammatory modulation.⁴² Similar findings were reported in studies in mice genetically predisposed to a severe protein C deficiency.⁴³ Interestingly, reconstitution of protein C levels in these mice with recombinant human activated protein C resulted in less severe inflammatory responses and an improved survival. In a model of severe abdominal infection through caecal ligation and puncture mice with a heterozygous deficiency of protein C had more profound organ dysfunction and an enhanced mortality in comparison with wild-type mice.⁴⁴ Taken together, these data suggest that preexistent protein C deficiency aggravates the coagulopathic response to severe infection and sepsis and is related to a worse outcome. It is not clear whether this observation may be extended to the clinical situation, mostly due to the fact that deficiency of protein C in humans is relatively rare. Therefore it is hard to establish a relationship between this condition and the incidence or outcome of sepsis.

FACTOR V LEIDEN IN MODELS OF EXPERIMENTAL INFECTION AND SEPSIS

In view of the central role of the protein C pathway in sepsis, a lot of attention has been given to the presence of factor V Leiden mutation, which leads to resistance to activated protein C, and the severity and outcome of sepsis or severe infection. In a clinical study in 259 children with meningococcal sepsis, factor V Leiden carriers had more profound coagulopathy and purpura fulminans, but their carrier status did not have a significant effect on survival.⁴⁵ Unfortunately, so far neither experimental or clinical studies in sepsis have shown unequivocal results regarding the presence of the factor V Leiden mutation. In one study, endotoxaemic mice carrying a heterozygous factor V Leiden mutation had a surprisingly lower mortality (19%) compared with their wild-type controls (57%).⁴⁶ In these experiments, factor V Leiden

mice produced more thrombin than normal controls, indicating a more profound activation of coagulation. In contrast, in another study of experimental pneumococcal pneumonia in mice no major protective effect of the factor V Leiden mutation was seen.⁴⁷ Also, markers of coagulation activation, both systemically and in the bronchoalveolar compartment, were not different between factor V Leiden mice and wild-type littermates. Remarkably, homozygosity for the factor V Leiden mutation protected against lethality in mice that were treated with ceftriaxone. Also, factor V Leiden mice did not differ significantly in their response compared with wild-type mice in a model of septic peritonitis, as reflected by similar degrees of activation of coagulation, inflammation, organ dysfunction and survival.⁴⁸ In another experimental study the effect of the presence of one or two alleles of the factor V Leiden mutation was investigated in lethal H1N1 influenza.⁴⁹ Factor V Leiden mutation did not influence the procoagulant response, lung histopathology, or survival in this study. Lastly, however, in a more subtle model of endotoxin-induced coagulation activation in humans it was demonstrated that heterozygous carriers of factor V Leiden had a more pronounced increase in markers of thrombin generation and fibrinogen to fibrin conversion.⁵⁰ The authors also found an increase in fibrinolytic activity in factor V Leiden affected individuals, which they attributed to the facilitating role of soluble fibrin for endogenous fibrinolysis.

Taken together, it seems that factor V Leiden may have some effect on the coagulopathy associated with sepsis; however, the effect is so subtle that it does not seem to be relevant against the background of the profound derangement of coagulation that is seen in severe sepsis or overwhelming infection.

FACTOR V LEIDEN IN CLINICAL SEPSIS STUDIES

Clinical studies on the role of factor V Leiden in sepsis also show variable results. The presence of the factor V Leiden mutation was analysed in large cohorts of patients with severe sepsis that had been included in intervention studies with recombinant human activated protein C.^{38,51} In this cohort of 3894 patients, the prevalence of factor V Leiden heterozygosity was 3.9%, which is slightly higher than the predicted allelic frequency of 2.5%.⁵² The 28-day mortality in those with factor V Leiden was not significantly different from the control population (19.3 vs 26.2%, respectively; risk ratio 0.74; 95% confidence interval (CI) 0.53-1.03). Moreover there were no differences in the incidence of serious bleeding or thrombotic events between factor V Leiden carriers and non-factor V Leiden carriers. In another publication in which the data of only

one of these two studies were presented, patients with a heterozygous factor V Leiden mutation were shown to have a lower mortality (13.9%) than those without this mutation (27.9%; $p=0.013$).⁴⁶ The effect of treatment with recombinant human activated protein C did not differ between the two groups. In the Copenhagen City Heart study 9253 individuals were screened for the presence of the factor V Leiden mutation and followed for a period of more than seven years to establish the risk of hospitalisation for any infectious disease and the subsequent risk of progression of disease to death.⁵³ The relative risk of any infection in carriers of the factor V Leiden mutation was 1.08 (95% CI 0.87-1.35) as compared with noncarriers (after adjustment for age, sex, smoking, alcohol consumption, income and level of education). In contrast with the previously mentioned study, patients with the factor V Leiden mutation in this study had a higher risk of death from infection as compared with patients who did not have this mutation (adjusted relative risk 4.41; 95% CI 1.42-13.67). The same group of authors presented data from four case cohorts of patients with either Gram-negative sepsis, or invasive pneumococcal disease, or intensive care admission.⁵⁴ When they compared their 1249 patients with matched controls, they found in an adjusted logistic regression analysis that factor V Leiden carriers had a higher risk of intensive care admission (OR 1.62; 95% CI 1.08-2.42) and were at increased risk of death (relative risk 1.78; 95% CI 1.13-2.81) compared with controls. Factor V Leiden was not associated with susceptibility to or outcome from pneumococcal infection or sepsis. Similarly, in a prospective observational study in 73 patients admitted with severe sepsis, the presence of a factor V Leiden mutation had no effect on short- or long-term mortality or any other clinically significant outcome.⁵⁵ Lastly, and in contrast with previous studies, a small study in 106 patients with acute respiratory distress syndrome showed a survival benefit in factor V Leiden heterozygotes (30-day survival 7/7 = 100% compared with 57/99 = 58% in patients without the mutation).⁵⁶ Obviously, this observation needs confirmation in larger cohorts of patients.

CONCLUSION

Activation of coagulation seems to play a pivotal role in the pathogenesis and outcome of severe infection and sepsis. Hypothetically, a preexisting prohaemostatic state, as seen in congenital thrombophilia, may aggravate the severity of this coagulopathy and may thereby affect outcome. Both experimental and clinical studies show inconsistent results as to a difference in survival from sepsis or severe infection in carriers of the factor V Leiden mutation. Although it may be biologically plausible that the factor V Leiden

mutation and the ensuing activated protein C resistance would aggravate the response to sepsis, the opposite may also be true as it has been speculated that a balanced and moderate increase in thrombin generation, as may be caused by a heterozygous factor V Leiden mutation, might be protective during severe infection and sepsis by means of generating slightly more activated protein C.⁵⁷ Additional analyses in larger cohorts of septic patients or long-term prospective studies in patients with a known factor V Leiden mutation will be required to clarify this issue.

REFERENCES

1. Levi M. Disseminated intravascular coagulation: a disease-specific approach. *Semin Thromb Hemost.* 2010;36:363-5.
2. Levi M, ten Cate H. Disseminated intravascular coagulation. *N Engl J Med.* 1999;341:586-92.
3. Levi M, Schultz M, van der Poll T. Disseminated intravascular coagulation in infectious disease. *Semin Thromb Hemost.* 2010;36:367-77.
4. Anas A, Wiersinga WJ, de Vos AF, van der Poll T. Recent insights into the pathogenesis of bacterial sepsis. *Neth J Med.* 2010;68:157-62.
5. Wheeler AP, Bernard GR. Treating patients with severe sepsis. *N Engl J Med.* 1999;340:207-14.
6. Levi M, de Jonge E, van der Poll T. Sepsis and disseminated intravascular coagulation. *J Thromb Thrombolysis.* 2003;16:43-7.
7. Levi M, Keller TT, van Gorp E, ten Cate H. Infection and inflammation and the coagulation system. *Cardiovasc Res.* 2003;60:26-39.
8. Levi M, van der Poll T. Inflammation and coagulation. *Crit Care Med.* 2010;38:S26-34.
9. Robboy SJ, Major MC, Colman RW, Minna JD. Pathology of disseminated intravascular coagulation (DIC). Analysis of 26 cases. *Hum Pathol.* 1972;3:327-43.
10. Shimamura K, Oka K, Nakazawa M, Kojima M. Distribution patterns of microthrombi in disseminated intravascular coagulation. *Arch Pathol Lab Med.* 1983;107:543-7.
11. Lowenberg EC, Meijers JC, Levi M. Platelet-vessel wall interaction in health and disease. *Neth J Med.* 2010;68:242-51.
12. Coalson JJ. Pathology of sepsis, septic shock, and multiple organ failure. Perspective on sepsis and septic shock. Fullerton, CA, Society of Critical Care Medicine, 1986, pp 27-59.
13. Levi M. The coagulant response in sepsis. *Clin Chest Med.* 2008;29:627-42, viii.
14. Creasey AA, Chang AC, Feigen L, Wun TC, Taylor FBJ, Hinshaw LB. Tissue factor pathway inhibitor reduces mortality from *Escherichia coli* septic shock. *J Clin Invest.* 1993;91:2850-6.
15. Kessler CM, Tang Z, Jacobs HM, Szymanski LM. The suprapharmacologic dosing of antithrombin concentrate for *Staphylococcus aureus*-induced disseminated intravascular coagulation in guinea pigs: substantial reduction in mortality and morbidity. *Blood.* 1997;89:4393-401.
16. Taylor FBJ, Chang A, Ruf W, et al. Lethal *E. coli* septic shock is prevented by blocking tissue factor with monoclonal antibody. *Circ Shock.* 1991;33:127-34.
17. Taylor FBJ, Chang A, Esmon CT, D'Angelo A, Vigano-D'Angelo S, Blick KE. Protein C prevents the coagulopathic and lethal effects of *Escherichia coli* infusion in the baboon. *J Clin Invest.* 1987;79:918-25.
18. Welty-Wolf KE, Carraway MS, Miller DL, et al. Coagulation blockade prevents sepsis-induced respiratory and renal failure in baboons. *Am J Respir Crit Care Med.* 2001;164:1988-96.

19. Miller DL, Welty-Wolf K, Carraway MS, et al. Extrinsic coagulation blockade attenuates lung injury and proinflammatory cytokine release after intratracheal lipopolysaccharide. *Am J Respir Cell Mol Biol.* 2002;26:650-8.
20. Fourrier F, Chopin C, Goudemand J, et al. Septic shock, multiple organ failure, and disseminated intravascular coagulation. Compared patterns of antithrombin III, protein C, and protein S deficiencies [see comments]. *Chest.* 1992;101:816-23.
21. Dhainaut JF, Yan SB, Joyce DE, et al. Treatment effects of drotrecogin alfa (activated) in patients with severe sepsis with or without overt disseminated intravascular coagulation. *J Thromb Haemost.* 2004;2:1924-33.
22. Levi M, Lowenberg EC. Thrombocytopenia in critically ill patients. *Semin Thromb Hemost.* 2008;34:417-24.
23. Vanderschueren S, De Weerd A, Malbrain M, et al. Thrombocytopenia and prognosis in intensive care. *Crit Care Med.* 2000;28:1871-6.
24. Levi MM, Eerenberg E, Lowenberg E, Kamphuisen PW. Bleeding in patients using new anticoagulants or antiplatelet agents: risk factors and management. *Neth J Med.* 2010;68:68-76.
25. Middeldorp S, Levi M. Thrombophilia: an update. *Semin Thromb Hemost.* 2007;33:563-72.
26. Inbal A, Kenet G, Zivelin A, et al. Purpura fulminans induced by disseminated intravascular coagulation following infection in 2 unrelated children with double heterozygosity for factor V Leiden and protein S deficiency. *Thromb Haemost.* 1997;77:1086-9.
27. Dogan Y, Aygun D, Yilmaz Y, et al. Severe protein S deficiency associated with heterozygous factor V Leiden mutation in a child with purpura fulminans. *Pediatr Hematol Oncol.* 2003;20:1-5.
28. al Ismail S, Collins P, Najib R, James-Ellison M, O'Hagan M. Postinfection purpura fulminans in a patient heterozygous for prothrombin G20210A and acquired protein S resistance. *Pediatr Hematol Oncol.* 1999;16:561-4.
29. Woods CR, Johnson CA. Varicella purpura fulminans associated with heterozygosity for factor V Leiden and transient protein S deficiency. *Pediatrics.* 1998;102:1208-10.
30. Sackesen C, Secmeer G, Gurgey A, et al. Homozygous Factor V Leiden mutation in a child with meningococcal purpura fulminans. *Pediatr Infect Dis. J.* 1998;17:87.
31. Levi M, Schouten M, van 't Veer C, van der Poll T. Factor V Leiden mutation in severe infection and sepsis. *Semin Thromb Hemost.* 2011; 37:955-60.
32. Texereau J, Pene F, Chiche JD, Rousseau C, Mira JP: Importance of hemostatic gene polymorphisms for susceptibility to and outcome of severe sepsis. *Crit Care Med.* 2004;32:S313-9.
33. Mesters RM, Mannucci PM, Coppola R, Keller T, Ostermann H, Kienast J. Factor VIIa and antithrombin III activity during severe sepsis and septic shock in neutropenic patients. *Blood.* 1996;88:881-6.
34. Levi M, Schouten M, van der Poll T. Sepsis, coagulation, and antithrombin: old lessons and new insights. *Semin Thromb Hemost.* 2008;34:742-6.
35. Minnema MC, Chang AC, Jansen PM, et al. Recombinant human antithrombin III improves survival and attenuates inflammatory responses in baboons lethally challenged with *Escherichia coli*. *Blood.* 2000;95:1117-23.
36. Yanada M, Kojima T, Ishiguro K, et al. Impact of antithrombin deficiency in thrombogenesis: lipopolysaccharide and stress-induced thrombus formation in heterozygous antithrombin-deficient mice. *Blood.* 2002;99:2455-8.
37. Levi M, de Jonge E, van der Poll T. Rationale for restoration of physiological anticoagulant pathways in patients with sepsis and disseminated intravascular coagulation. *Crit Care Med.* 2001;29(7 Suppl):S90-429:S90-S94.
38. Bernard GR, Vincent JL, Laterre PF, et al. Efficacy and safety of recombinant human activated protein C for severe sepsis. *N Engl J Med.* 2001; 344:699-709.
39. Lay AJ, Liang Z, Rosen ED, Castellino FJ. Mice with a severe deficiency in protein C display prothrombotic and proinflammatory phenotypes and compromised maternal reproductive capabilities. *J Clin Invest.* 2005;115:1552-61.
40. Jalbert LR, Rosen ED, Moons L, et al. Inactivation of the gene for anticoagulant protein C causes lethal perinatal consumptive coagulopathy in mice. *J Clin Invest.* 1998;102:1481-8.
41. Levi M, Dorffler-Melly J, Reitsma PH, et al. Aggravation of endotoxin-induced disseminated intravascular coagulation and cytokine activation in heterozygous protein C deficient mice. *Blood.* 2003;101:4823-7.
42. Levi M, van der Poll T, Buller HR. Bidirectional relation between inflammation and coagulation. *Circulation.* 2004;109:2698-704.
43. Lay AJ, Donahue D, Tsai MJ, Castellino FJ. Acute inflammation is exacerbated in mice genetically predisposed to a severe protein C deficiency. *Blood.* 2007;109:1984-91.
44. Ganopoulos JG, Castellino FJ. A protein C deficiency exacerbates inflammatory and hypotensive responses in mice during polymicrobial sepsis in a cecal ligation and puncture model. *Am J Pathol.* 2004;165:1433-46.
45. Kondaveeti S, Hibberd ML, Booy R, Nadel S, Levin M. Effect of the Factor V Leiden mutation on the severity of meningococcal disease. *Pediatr Infect Dis J.* 1999;18:893-6.
46. Kerlin BA, Yan SB, Isermann BH, et al. Survival advantage associated with heterozygous factor V Leiden mutation in patients with severe sepsis and in mouse endotoxemia. *Blood.* 2003;102:3085-92.
47. Schouten M, van 't Veer C, Roelofs JJ, Levi M, van der Poll T. Impact of the factor V Leiden mutation on the outcome of pneumococcal pneumonia: a controlled laboratory study. *Crit Care.* 2010;14:R145.
48. Bruggemann LW, Schoenmakers SH, Groot AP, Reitsma PH, Spek CA. Role of the factor V Leiden mutation in septic peritonitis assessed in factor V Leiden transgenic mice. *Crit Care Med.* 2006;34:2201-6.
49. Schouten M, van der Sluijs KF, Roelofs JJ, Levi M, van 't Veer C, van der Poll T. Factor V Leiden mutation does not affect coagulopathy or outcome in lethal H1N1 influenza. *Eur Respir J.* 2010; 36:1346-54.
50. Elmas E, Suvajac N, Jilma B, Weiler H, Borggrefe M, Dempfle CE. Factor V Leiden mutation enhances fibrin formation and dissolution in vivo in a human endotoxemia model. *Blood.* 2010;116:801-5.
51. Bernard GR, Margolis BD, Shanies HM, et al. Extended evaluation of recombinant human activated protein C United States Trial (ENHANCE US): a single-arm, phase 3B, multicenter study of drotrecogin alfa (activated) in severe sepsis. *Chest.* 2004;125:2206-16.
52. Yan SB, Nelson DR. Effect of factor V Leiden polymorphism in severe sepsis and on treatment with recombinant human activated protein C. *Crit Care Med.* 2004;32:S239-46.
53. Benfield TL, Dahl M, Nordestgaard BG, Tybjaerg-Hansen A. Influence of the factor V Leiden mutation on infectious disease susceptibility and outcome: a population-based study. *J Infect Dis.* 2005;192:1851-7.
54. Benfield T, Ejrnaes K, Juul K, et al. Influence of Factor V Leiden on susceptibility to and outcome from critical illness: a genetic association study. *Crit Care.* 2010;14:R28.
55. Tsantes AE, Tsangaris I, Bonovas S, et al. The effect of four hemostatic gene polymorphisms on the outcome of septic critically ill patients. *Blood Coagul Fibrinolysis.* 2010;21:175-81.
56. Adamzik M, Frey UH, Riemann K, et al. Factor V Leiden mutation is associated with improved 30-day survival in patients with acute respiratory distress syndrome. *Crit Care Med.* 2008;36:1776-9.
57. Weiler H, Kerlin B, Lytle MC. Factor V Leiden polymorphism modifies sepsis outcome: evidence from animal studies. *Crit Care Med.* 2004;32:S233-8.